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REMARKS

Upon the entry of this preliminary amendment, claims 101-132 will be pending in this application. Claims 101-132 (elected Group XXVII) relate to GPR38(V297K), and claims 133-144 relate to the use of GPR38(V297K). The Office Action indicates that claims 101-132 are product claims, and claims 133-144 are process claims. Further, the Office Action indicates that the process claims 133-144 should be withdrawn, and may be rejoined later when the product claims 101-132 are allowed. See the Advisory Action at page 3, last paragraph. For the sole purpose of facilitating prosecution, Applicants hereby withdraw claims 133-144, but nevertheless maintain the right to rejoin these claims when claims 101-132 are allowed.

In a preliminary matter, Applicants note that the arguments set forth by the Office Action mailed December 17, 2003 and the Advisory Action mailed March 9, 2004 are substantially the same. Thus, the Office Action and the Advisory Action will be referred to herein as the "Office Action", unless otherwise specified.

Claims 101-132 are rejected under 35 U.S.C. §101 by the Office Action for allegedly lacking a "well-established utility." Applicants respectfully disagree with the Office Action, and assert that the claimed inventions have a well-established utility.

According to the MPEP §2107.02 II.B, an invention has a well-established utility if:

- (i) a person of ordinary skill in the art would immediately appreciate why the invention is useful based on the characteristics of the invention, and
- (ii) the utility is specific, substantial and credible.

It would have been readily apparent to one of ordinary skill in the art that the claimed inventions directed to GPR38(V297K) would have a well established utility.

A. Those of Skill in the Art Would Have Immediately Appreciated Why the Invention is Useful

On the priority filing date of the present application (March 12, 1999), a person of

ordinary skill in the art would have immediately appreciated why the invention is useful. Applicants submit herewith the Declaration of Dr. Dominic P. Behan (dated June 7, 2004), in which he states that on March 12, 1999, one of ordinary skill in the art (e.g., biochemists and molecular biologists) would have immediately appreciated that the claimed invention directed to the receptor GPR38(V297K) would have been useful. As Dr. Behan explains, one of ordinary skill would have immediately appreciated that an invention directed to a GPR38(V297K) receptor would have been useful, because the receptor GPR38(V297K) could have been used in an assay to identify an inverse agonist of the receptor GPR38, wherein such inverse agonists could have been employed to prevent the exacerbation of, or to treat, Graves' disease. Specifically, in paragraph 5 of the Declaration, Dr. Behan states that as of March 12, 1999, biochemists and molecular biologists would have immediately appreciated that the claimed invention directed to the receptor GPR38(V297K) would have been useful because:

- (a) the non-endogenous GPR38(V297K) differs from the endogenous GPR38 by a single amino acid, and yields a constitutively active version of the endogenous GPR38;
- (b) the constitutively active GPR38(V297K) causes increased production of intracellular cAMP (see Example 4 and Figure 1 of U.S. Provisional Application Ser. No. 60/123,945; enclosed herein as Exhibits 2 and 3, respectively);
- (c) GPR38 is expressed in the thyroid;
- (d) an activated GPR38 is functionally similar to a GPR38(V297K);
- (e) therefore, an activated GPR38 in the thyroid would cause an increase in production of intracellular cAMP therein;
- (f) an elevated level of intracellular cAMP in the thyroid leads to an over production of thyroid hormones in Graves' disease (see Goodman and Gilman's The Pharmacological Basis of Therapeutics, McGraw-Hill, Ninth Edition (1996) pages 1383-1409, previously submitted in the Response dated February 12, 2004 as Exhibit A; enclosed herein as Exhibit 4); and

- (g) GPR38(V297K) could have been used in an assay to identify an inverse agonist of GPR38;
- (h) such an inverse agonist would have decreased the level of cAMP in the thyroid, and therefore would have been useful to decrease the production of thyroid hormones to prevent the exacerbation of, or to treat, Graves' disease.

Thus, a person of ordinary skill in the art would have immediately appreciated that the claims directed to GPR38(V297K) would have at least one well-established utility because a GPR38(V297K) could have been employed in screening assays to identify, for example, inverse agonists that could have been used to prevent the exacerbation of, or for the treatment of, Graves' disease.

B. Applicants' Asserted Utility is Specific, Substantial and Credible

1. Applicants' Asserted Utility is Specific

The utility would have been specific because, for example, it was known that GPR38 is specifically expressed in the thyroid, and the inhibition of an activated GPR38 (e.g., by an inverse agonist identified through a screening assay that employs GPR38(V297K)) would have led to a decreased production of cAMP specifically in the thyroid, which in turn could have been used specifically to prevent the exacerbation of, or for the treatment of, Graves' disease. See Dr. Behan's Declaration at paragraph 5.

2. Applicants' Asserted Utility is Substantial

Applicants' asserted utility would have been substantial because, for example, preventing the exacerbation of, or treatment of, Graves' disease would have been a "real world" use. More specifically, the use of GPR38(V297K) in an assay to identify possible inverse agonists thereof, wherein the inverse agonists could have been used to prevent the exacerbation of, or for the treatment of, Graves' disease would have been a "real world" use. In this regard, Applicants note that the Revised Interim Utility Guidelines Training Material (herein after "Training

Material") states that "an assay method for identifying compounds that themselves have a 'substantial utility' define a 'real world' context of use." See page 6 of the Training Material, which was submitted as Exhibit B with the Response filed on February 12, 2004. In the present case, the compounds that could have been identified in an assay employing GPR38(V297K) would have had substantial utility themselves because these compounds, e.g. inverse agonists of GPR38, could have been administered to prevent the exacerbation of, or for the treatment of, Graves' disease. Thus, the use of GPR38(V297K) in an assay to identify possible inverse agonists thereof also would have been a "real world" use.

3. Applicants' Asserted Utility is Credible

As Dr. Behan states in his Declaration, the asserted utility would have been credible because the use of GPR38(V297K) to screen for inverse agonists compounds, wherein such compounds could have been administered to a patient to prevent the exacerbation of, or for the treatment of Graves' disease, would have been believable to a person of ordinary skill in the art. See paragraph 6 of Dr. Behan's Declaration.

4. The Office Has Not Provided Any Evidence that Those of Skill in the Art Would Have Found the Asserted Utility to be Not Credible

The MPEP dictates that once the applicants have provided a reason for why the claimed invention is useful, the Office personnel may maintain a rejection for alleged lack of utility only if the Office Action establishes that one of ordinary skill would find that the asserted utility is not credible. For example, the MPEP §2107.02 II. B. states that:

If the applicant subsequently indicates why the invention is useful, Office personnel should review that assertion according to the standards articulated below for review of the credibility of an asserted utility.

The "standard articulated below for review of the credibility of an asserted utility" is found at MPEP §2107.02 III. B, which states:

Where an applicant has specifically asserted that an invention has a particular utility, that assertion cannot simply be dismissed by Office personnel as being

"wrong," even when there may be reason to believe that the assertion is not entirely accurate. Rather, Office personnel must determine if the assertion of utility is <u>credible</u> (i.e., whether the assertion of utility is believable to a person of ordinary skill in the art based on the totality of evidence and reasoning provided). An assertion is credible unless (A) the logic underlying the assertion is seriously flawed, or (B) the facts upon which the assertion is based are inconsistent with the logic underlying the assertion. Credibility as used in this context refers to the reliability of the statement based on the logic and facts that are offered by the applicant to support the assertion of utility.

(emphasis in original).

Dr. Behan states in his Declaration that the use of GPR(V297K) to screen for inverse agonists of GPR38, wherein such inverse agonists could have been administered to prevent the exacerbation of, or for the treatment of, Graves' disease, would have been believable to a person of ordinary skill in the art on March 12, 1999 (the earliest priority date of the present application). See paragraph 6 of Dr. Behan's Declaration. There is no evidence of record that would contradict the logic underlying Dr. Behan's assertion, as set forth in his Declaration, or that would indicate that the facts upon which Dr. Behan's assertion is based are inconsistent with the logic underlying the assertion. Indeed, should the Office question any of the statements of fact in Dr. Behan's Declaration, Applicants would then request that the Office provide evidence to rebut these statements. See *In re Alton*, 76 F.3d 1168 (Fed. Cir. 1996), and The Federal Register, Vol. 66, No. 4, January 5, 2001, pages 1098-1099.

C. The Arguments of the Office Action Have Been Overcome by the Behan Declaration

The Office Action alleges that the invention lacks utility on the basis that the normal physiological role and endogenous ligand for GPR38 (and therefore GPR38(V297K) is unknown. See the Office Action of December 17, 2003, at pages 5-6. However, the knowledge of normal physiological role and endogenous ligand for GPR38 is irrelevant to determining whether the asserted well-established utility for GPR38(V297K) is credible. Rather, the relevant inquiry is whether the following asserted facts recited by Dr. Behan in his Declaration are credible: (1) at the time of filing of the present application, GPR38 is known to be in specific

locations, e.g., the thyroid; (2) an activated GPR38 would promote the production of cAMP in the thyroid; (3) patients suffering from Graves' disease would benefit from the reduction of cAMP in the thyroid; and (4) GPR38(V297K) could have been used to screen for inverse agonists that could have been administered to act on the endogenous GPR38 to reduce the thyroid production of cAMP. See Dr. Behan's Declaration at paragraph 5. Inasmuch as the Office Action has not provided any evidence that those of ordinary skill would doubt the credibility of these asserted facts, the present invention must be deemed to have the asserted well-established utility.

The Office Action asserts that there is "no disclosure that changing cAMP levels in the thyroid by modulating GPR38(V297K)¹ or GPR38 would have any effect on Graves' disease." Office Action of December 17, 2003 at page 6. However, it is not necessary that such an assertion be present in the specification. In this regard, Applicants note that either an explicitly stated utility, <u>or</u> an asserted specific and substantial utility is sufficient to satisfy 35 USC § 101. Indeed, the title of MPEP § 2107.02 II.B itself reads: "No statement of utility for the claimed invention in the specification does not per se negate utility". Thus, Applicants' assertion that the claimed invention has a well-established utility is sufficient to meet the requirements of 35 USC § 101.²

The Office Action states that the specification discloses general functional activities of G-protein coupled receptors (GCPR), but does not disclose any activity associated with the specific GPR38(V297) of the instant invention. Office Action of December 17, 2003 at page 17. However, contrary to the Office Action's allegation, the utility of GPR38(V297K) would have been specific because it is known that GPR38 is specifically found in the thyroid, and that the inhibition of an activated GPR38 (e.g., by an inverse agonist identified through a screening assay

Applicants note the GPR38(V297K) is not endogenous. Thus, any consideration as to whether modulation of GPR38(V297K) would have any effect on Graves' disease would be irrelevant as to the present utility analysis. The well-established utility for GPR38(V297K) is that it could have been used in a screening assay to identify compounds that may modulate GPR38, for example an endogenous GPR38.

² To the extent that the Office Action intends to question the veracity of Applicants' assertion regarding the effect of changing cAMP levels on Graves Disease, Applicants respectfully assert that the burden is on the Office to provide reasons why those of skill in the art would question Dr. Behan's statement.

that employs GPR38(V297K)) would have led to a decreased production of cAMP specifically in the thyroid, which in turn would have been specifically useful to prevent the exacerbation of, or for the treatment of, Graves' disease. See Dr. Behan's Declaration at paragraph 5.

The Office Action further states that the "asserted utilities are essentially methods of testing for or for potentially treating unspecified, undisclosed diseases or conditions, which does not define a 'real world' context of use." Office Action of December 17, 2003 at page 8. However, contrary to the Office Action's allegation, an inverse agonist that would have been identified by GPR38(V297K) could have been administered to prevent the exacerbation of, or for the treatment of, Graves' disease, as discussed above and in Dr. Behan's Declaration. Thus, the asserted utility does have a "real world" use.

The Office Action further states that GPR38(V297K) is a "new member" to the GCPR family, wherein the existing members have divergent functions, and further states that "without some common biological activity for the family members, a new member would not have a specific, substantial or credible utility." Office Action of December 17, 2003 at pages 9-10. However, whether GPR38(V297K) is a "new member" of the GPCR family, and whether an activity is known to be common to all members of the GPCR family, is irrelevant to whether GPR38(V297K) has a well-established utility. Rather, the proper inquiry is whether GPR38(V297K) is useful, for example, in identifying inverse agonists that could have been employed to prevent the exacerbation of, or for the treatment of, Graves' disease, as described in Dr, Behan's Declaration. Inasmuch as the Office Action has failed to show why GPR38(V297K) would not have such use, the present claimed invention must be deemed to have the asserted well-established utility.

The Office Action further asserts that:

... any compound could be considered a regulator or modulator of tissue in that any compound, if administered in the proper amount, will stimulate or inhibit tissue. For example, salt, ethanol, and water area all compounds which kill cells if administered in a great enough amount, and which would stimulate cells from which these compounds had been withheld, therefore, they could be considered regulators or modulators of tissue.

PATENT

AREN-0240 (AREN-7.US27.DIV)

Office Action of December 17, 2003 ay page 10. However, as discussed above and in Dr.

Behan's Declaration, GPR38(V297K) could have been employed in a screening assay to identify

compounds, for example inverse agonists, that can inhibit the activity of a specific receptor

(GPR38) that is localized in a specific tissue (the thyroid). Thus, the comparison proposed by

the Office Action is completely inappropriate.

In view of the preceding remarks, Applicants respectfully request the rejections under 35

U.S.C. § 101 be withdrawn.

Claims 101-132 also stand rejected under 35 U.S.C. § 112, first paragraph, for alleged

lack of enablement, on the basis that the claims allegedly lack utility. In light of the arguments

above, Applicants respectfully submit that those skilled in the art would recognize both the

utility of the invention and how to use it. Applicants therefore respectfully request withdrawal of

the rejection under 35 U.S.C. § 112, first paragraph.

In conclusion, Applicants respectfully assert that the claimed inventions directed to

GPR38(V297K) have a well-established utility, and that the Office Action has not provided any

reason for one of ordinary skill in the art to doubt the credibility of such utility. Accordingly,

Applicants respectfully request a withdrawal of the rejection under 35 U.S.C. § 101 and § 112,

first paragraph. Further, Applicants assert that the claims are in condition for allowance, and

respectfully request notification to that effect. Should the Office have any questions, Applicants

invite the Office to contact the undersigned at (215) 665-2158 to discuss any issues unresolved

by this Amendment. A Notice of Allowance is earnestly solicited.

Respectfully submitted,

Date: June 17, 2004

Quan L. Nguyen

Reg. No. 46,957

COZEN O'CONNOR, P.C.

1900 Market Street

Philadelphia, PA 19103-3508

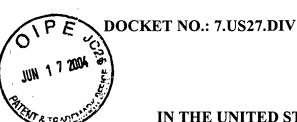
Telephone: 215.665.2158

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Enclosures:

Doc No. 2084033

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of: Examiner:

Behan et al. N. Basi

Serial No.: 09/876,252 Group Art Unit: 1646

Filed: June 7, 2001 Confirmation No. 8181

For: NON-ENDOGENOUS CONSTITUTIVELY ACTIVATED HUMAN G

PROTEIN-COUPLED RECEPTORS

DECLARATION PURSUANT TO 37 C.F.R. §1.132

- I, Dominic P. Behan, Ph.D., do hereby declare as follows:
- 1. I am the co-founder and Vice President of Research at Arena Pharmaceuticals, Inc. A copy of my *curriculum vitae* is enclosed herein as Exhibit 1.
- 2. I am an inventor of the above referenced patent application. I am familiar with the level of skill of those in the art of biochemistry and molecular biology on March 12, 1999, which I am informed is the earliest priority date of the above referenced patent application.

DOCKET NO.: 7.US27.DIV PATENT

3. I understand that the pending claims relate to a non-endogenous G-protein-coupled receptor, GPR38(V297K).

d

- 4. I have read the Advisory Action and the Final Office Action from the Patent Office dated March 9, 2004 and December 17, 2003, respectively. I understand that the Office Action alleges that the pending claims lack utility.
- 5. I believe that the allegation by the Office Action is incorrect. As of March 12, 1999, biochemists and molecular biologists would have immediately appreciated that the claimed invention directed to the receptor GPR38(V297K) would have been useful because:
 - (a) the non-endogenous GPR38(V297K) differs from the endogenous GPR38 by a single amino acid, and yields a constitutively active version of the endogenous GPR38;
 - (b) the constitutively active GPR38(V297K) causes increased production of intracellular cAMP (see Example 4 and Figure 1 of U.S. Provisional Application Ser. No. 60/123,945; enclosed herein as Exhibits 2 and 3, respectively);
 - (c) GPR38 is expressed in the thyroid;
 - (d) an activated GPR38 is functionally similar to a GPR38(V297K);
 - (e) therefore, an activated GPR38 in the thyroid would cause an increase in production of intracellular cAMP therein;
 - (f) an elevated level of intracellular cAMP in the thyroid leads to an over production of thyroid hormones in Graves' disease (see Goodman and Gilman's The Pharmacological Basis of Therapeutics, McGraw-Hill, Ninth

DOCKET NO.: 7.US27.DIV PATENT

Edition (1996) pages 1383-1409, previously submitted in the Response dated February 12, 2004 as Exhibit A; enclosed herein as Exhibit 4); and

(g) GPR38(V297K) could have been used in an assay to identify an inverse

agonist of GPR38;

eţ.

(h) such an inverse agonist would have decreased the level of cAMP in the

thyroid, and therefore would have been useful to decrease the production of

thyroid hormones to prevent the exacerbation of, or to treat, Graves' disease.

6. Moreover, the use of GPR38(V297K) to screen for inverse agonists of GPR38,

wherein such inverse agonists may be administered for the prevention of exacerbation of,

or for the treatment of, Graves' disease, as stated above, would have been believable to a

person of ordinary skill in the art on March 12, 1999.

7. I declare that statements made herein of my own knowledge are true and that all

statements made on information and belief to be true; and further that these statements

were made with knowledge that willful false statements and the like so made are

punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United

States Code that such willful false statements may jeopardize the validity of the

application or any patent issued thereon.

Dated: 6-7-04

Dominic Behan, Ph.D.

Doc No. 2074526

CURRICULUM VITAE

DOMINIC PHILIP BEHAN

DATE OF BIRTH: November 2, 1963

PLACE OF BIRTH: Liverpool, England

MARITAL STATUS: Married

Gaynor Behan

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(see above)

CITIZENSHIP: USA

CAREER SUMMARY

Dr. Dominic P. Behan obtained a BSc. Honors degree in Biochemistry from the University of Leeds, UK followed by a Ph.D. in Biochemistry from the University of Reading, UK working on corticotrophin releasing factor binding proteins (supervisor Prof. Phil Lowry). He then spent three years as a Post Doctoral Research Fellow in Prof. Wylie Vale's laboratory (Salk Institute, San Diego, CA) continuing to work on CRF and CRF binding proteins. His industrial career began at Neurocrine Biosciences, San Diego where he directed a number of programs from 1993-1997. The early part of his academic and industrial career has focused on identifying novel mechanisms for the inactivation of neuropeptides at peptide/neurotransmitter receptors in the periphery and central nervous system. This was mainly applied to corticotropin releasing factor (CRF), a 41 residue hypothalamic peptide which mediates the body's endocrine, behavioral, autonomic and immune responses to stress. His PhD and post doctoral work resulted in the isolation and molecular characterization of a novel binding protein (CRF-BP), a 37 KDa protein which inactivates the biological activity of all members of the CRF family of neuropeptides. This work established this factor as an important modulator of the hypothalamic-pituitary-adrenal (HPA) axis, the placental-HPA axis and the central actions of CRF and CRF related peptides such as urocortin. Furthermore, this work established a novel mechanism for the synaptic inhibition of these neuropeptides in human brain and resulted in the identification of a novel therapeutic drug target for the potential treatment of eating disorders, psychiatric illnesses and neurodegenerative diseases such as Alzheimer's disease. Dr. Behan also expanded this work and

published a mechanism for elevating "free" IGF-I levels by the use or orally active small molecules that displaced bound insulin-like growth factor (IGF) from its binding protein and resulted in increases in endogenous "free" levels of IGFs in blood and relevant brain areas. Dr. Behan was responsible for running two major programs at Neurocrine Biosciences which led to the consolidation of a major corporate alliance with Eli Lilly (announced 10/21/96) valued up to \$74 million. In 1997 he co-founded Arena Pharmaceuticals and has served as Vice President and Director since then. Dr. Behan has been involved in all aspects of Arena's business and scientific operations including establishment and management of the scientific infrastructure, management of corporate collaborations and fund raising through private equity and public offerings. In April 2000 he was elected to the board of Directors of Arena Pharmaceuticals. He is also co-inventor of Arena's CARTTM technology that has allowed for the screening of multiple G-protein coupled receptors (GPCRs) for small molecule ligands without knowledge of the natural ligand. His research interests include all aspects of drug discovery from receptors, particularly GPCRs. The current focus of this research has been to globally understand how GPCRs regulate biological systems that impact human disease and more importantly to identify small molecule drug candidates to this important class of protein.

EDUCATION & PREVIOUS POSITIONS

Bsc Hons Biochemistry (received June 1986)

University of Leeds, England

PhD in biochemistry (received February 1990) University of Reading, England

Post-Doctoral Research (1990-1993)

The Salk Institute for Biological

Studies, San Diego, USA

Ph.D. RESEARCH

The program began with the identification of a soluble binding protein for corticotropin-releasing-factor (CRF) in human plasma. The protein was then purified from human plasma within eighteen months which allowed time to complete a six month sequencing and cloning study (with Professor Wylie Vale, Salk Institute, CA, U.S.A; British MRC grant). The amino acid sequence of tryptic fragments of the protein was determined after HPLC purification. The first cDNA fragment was isolated by PCR amplification from a human liver and rat brain cDNA library. In addition to his research activities he also had some tutorial and supervisory responsibilities for undergraduates.

POST-DOCTORAL EXPERIENCE

As a continuation from the Ph.D. the research focused on the neuroendocrine regulation of stress and the synaptic regulation of CRF-like neuropetides in brain. Further, the transcriptional regulation of CRF-BP expression was investigated and protein isoforms of CRF-BP where characterized utilizing a variety of protein chemical and molecular biological techniques. The research on CRF-BP was expanded and resulted in the cloning and characterization of the human gene (promoter elements and the multi intronic/exonic structure were elucidated). In addition, the further purification, cloning and biochemical characterization of two membrane associated forms

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of CRF-BP from brain which are involved in the synaptic modulation of the CRF family of neuropeptides at neurotransmitter receptors was accomplished. Using deletion analysis, the work identified key peptide regions of CRF which bind to CRF-BP but do not interact with CRF and/or urocortin receptors. These would later prove very important for the validation of CRF-BP as a novel drug target for the treatment of Alzheimer's disease and obesity. In addition, the neuroanatomical localization of CRF-BP was determined in the CNS and correlated with CRF expression to understand how these factors may be regulating brain CRF containing neurons.

TEACHING EXPERIENCE & LECTURES/CHAIRS (1988-1996)

1988-1990	Demonstrator-Department of Biochemistry & Physiology Reading University, England.
1990-1993	Lecture/coordinator-UCSD graduate course in molecular biology & biochemistry, The Salk Institute For Biological Studies, San Diego.
1993-1997	Neurocrine monthly seminar series, Neurocrine Biosciences Inc. San Diego.
1994	Chairman at the 76th Annual meeting of the Endocrine Society.
1996	UCSD, School of Medicine, Department of Reproductive Medicine, Invited Speaker, November 1996.
1996	British Neuroendocrine Group. Annual Meeting. 2nd-3rd July, Edinburgh (invited symposium speaker).

PROFESSIONAL EXPERIENCE

Neurocrine Biosciences Inc., San Diego Group Leader (1993-January 1997)

- The work on CRF-BP was one of the founding concepts of Neurocrine Biosciences Inc. Dr. Behan was one of the initial founding scientists.
- Company was mainly founded on drug discovery around the corticotropinreleasing factor system. Dr. Behan's work on corticotropin releasing factorbinding protein (CRF-BP) established this as a major drug target and a key Neurocrine program for launching the company.
- Established presence of CRF-BP in control and Alzheimer's human brain.
- Developed novel technology to measure bound and "free" levels of this and other neuropeptides.

- Established proof of concept for the utility of ligand inhibitors of CRF-BP for the treatment of Alzheimer's disease and obesity resulting in a publication in Nature and Proceedings of the National Academy of Sciences (USA) (see publications).
- Played a major role in presentations and R&D negotiations with potential cooperate partners to consolidate a major corporate collaboration with Eli. Lilly valued up to \$75 million.
- Identified a new technology platform with multiple novel drug targets for diabetes, obesity and neuronal degeneration/ischemia and established a second fully staffed program.

Arena Pharmaceuticals, Inc

Co-founder and Vice President of Research (January 1997-present)

In 1997 he co-founded Arena Pharmaceuticals and has served as Vice President and Research Director since then. In April 2000 he was elected to the board of Directors of Arena Pharmaceuticals. He is also co-inventor of Arena's CARTTM technology. His research interests include all aspects of drug discovery from receptors, particularly GPCRs.

- Co-founder
- Participated in the raising of over \$230 million in private and public equity
- Established alliances Eli Lilly, Taisho, Fujisawa, Astra Zeneca, ICI, Tripos and Neurocrine Biosciences Inc.
- Responsible for overseeing Arena's research of > 300 FTE
- Author of > 100 patent applications on GPCRs and CART technology
- Elected to Board of Directors, April 2000

Reviewer of scientific journals

Dr. Behan been requested to review numerous scientific journals and publications.

CORPORATE COLLABORATIONS

Dr. Behan has been responsible and/or involved in coordinating the following alliances at Arena Pharmaceuticals Inc.

Eli Lilly Taisho Fujisawa Neurocrine Biosciences Astra Zeneca

AWARDS & HONORS

1987-1990

British Medical Research Council Scholarship

1989-1990	British Medical Research Council Visitor Exchange Scholarship (this funded a six month trip to the Salk Institute, San Diego during my PhD to sequence the purified corticotropin-releasing factor-binding protein)
1990-1993	Adler Foundation Scholarship (funded a 3 year post-doctorate at the Salk Institute, San Diego)
1994-1995	Small Business Phase I Innovation Grant from the department of health and human services (this grant was obtained to study the importance of the binding protein and CRF in Alzheimer's disease)
1995-1996	Small Business Phase II Innovation Grant from the department of health and human services (CRF and Alzheimer's disease)
1996-Present	Small business phase I innovation grant to study a novel therapeutic drug target for diabetes

AFFILIATIONS (Past & present)

Society for Neuroscience
Endocrine Society
New York Academy of Sciences
American Association for the Advancement of Science
Winter Neuropeptide Group
British Neuroendocrine Group (BNG)

PUBLICATIONS

Papers & Review Articles

- 1. Linton, E.A., Wolfe, C.D.A., **Behan, D.P.** and Lowry, P.J. (1988). A specific carrier substance for human corticotropin releasing factor in late gestational maternal plasma which could be masking it's ACTH-releasing activity. Clinical Endocrinology, **28**, 315, 324.
- 2. **Behan, D.P.**, Linton, E.A. and Lowry, P.J. (1989). Isolation of the human plasma corticotropin releasing factor binding protein. Journal of Endocrinology, **121**, 23-31.
- 3. Linton, E.A., **Behan, D.P.** and Lowry, P.J. (1989). The CRF binding protein in human plasma. In Control of the hypothalamo-pituitary-adrenocortical axis. (ed) FC Rose. International universities press, Madison, pp 25-36.
- 4. Linton, E.A., **Behan**, **D.P.**, Saphier, P.W. and Lowry, P. J. (1989). Corticotropin-Releasing Hormone Binding protein: Reduction in the ACTH-releasing activity of

- placental but not hypothalamic CRH. J. Clin. Endocr. Metab., 70, 1574-1580.
- 5. Potter, E., **Behan, D.P.**, Fischer, W., Linton, E.A., Lowry, P.J. and Vale, W. (1991). Cloning and characterization of the cDNAs for the human and rat corticotropin releasing factor binding proteins. Nature, **349**, 423-426.
- 6. Potter, E., **Behan, D.P.,** Linton, E.A., Lowry, P.J. Sawchenko, P.E. and Vale, W. (1991). The Central Distribution of a CRF-Binding Protein Predicts Multiple Sites and Modes of Interaction with CRF. Proc. Natl. Acad. Sci. (USA), **89**, 423-426.
- 7. Linton, E.A., Perkins, A.V., Woods, R.J., Eben, F., Wolfe, C.D.A., **Behan, D.P.**, Potter, E., Vale, W.W. and Lowry, P.J. (1992). Corticotropin releasing hormone-binding protein [CRH-BP]: plasma levels decrease during the third trimester of normal human pregnancy. J. Clin. Endocrinol. Metab., **76**, 260-262.
- 8. **Behan, D.P.**, Potter, E., Linton, E.A., Lowry, P.J. and Vale, W. (1993). Cloning and structure of the human CRF-binding protein gene. Genomics, 16, 63-68.
- 9. **Behan, D.P.**, Potter, E.A., Sutton, S., Fischer, W., Lowry, P.J. and Vale, W.W. (1993). Corticotrophin-releasing factor-binding protein: A putative peripheral and central modulator of the CRF family of neuropeptides. Ann. N.Y. Acad. Sci., **697**, 1-8.
- 10. Petraglia, F., Potter, E., Cameron, V.A., Sutton, S., **Behan, D.P.**, Woods, R.J., Sawchenko, P.E., Lowry, P.J. and Vale, W. (1993). Corticotropin-releasing factor binding protein is produced by human placenta and intrauterine tissues. J. Clin. Endocrinol. Metab., **77**, 919-923.
- 11. Fischer, W.H., **Behan, D.P.**, Park, M., Potter, E., Lowry, P.J. and Vale, W. (1994). Assignment of disulfide bonds in corticotropin releasing factor-binding protein. J. Biological Chem., **6**, 4313-4316.
- 12. Woods, R.J., Grossman, P., Saphier, P., Kennedy, K., Ur, E., **Behan, D.P**, Potter, E., Vale, W. and Lowry, P.J. (1994). Association of human corticotropin-releasing hormone to its binding protein in blood may trigger clearance of the complex. J. Clin. Endocrinol. Metab., 78, 73-76.
- Woods, R.J., Kennedy, K.M., Gibbins, J.M., **Behan, D.P.**, Vale, W. and Lowry, P.J. (1994). Corticotropin-releasing factor binding protein dimerizes after association with ligand. Endocrinology, **135**, 768-773.
- 14. **Behan, D.P.**, Cepoi, D., Fisher, W.H., Park, M., Lowry, P.J., Sutton, S. and Vale W.W. (1995). Characterization of a sheep brain corticotropin releasing factor binding protein. Brain Res., 709, 265-274.
- 15. **Behan, D.P.**, De Souza, E.B., Lowry, P.J., Potter, E., Sawchenko, P. and Vale, W. (1995). Corticotropin releasing factor (CRF) binding protein: a novel regulator of CRF

- and related peptides. Frontiers In Neuroendocrinology, 16, 362-382.
- 16. **Behan, D.P.**, Heinrichs, S.C., Troncoso, J.C., Liu, X., Kawas, C.H., Ling, N. and De Souza, E. (1995). Corticotropin releasing factor displacement from its binding protein as a possible treatment for Alzheimer's disease. Nature, **378**, 284-287.
- 17. **Behan, D.P.**, Khongsaly, O., Liu, X., Ling, N., Goland, R., Nasman, B., Olsson, T., Troncoso, J.C. and De Souza, E.B. (1995). Measurement of corticotropin releasing factor (CRF) binding protein (CRF-BP) and CRF/CRF-BP complex in human plasma by two-site enzyme linked immunoabsorbant assay. J. Clin. Endocr. Metab., **81**, 2579-2586.
- 18. **Behan, D.P.**, Maciejewski, D., Chalmers, D. and De Souza, E.B. (1995). Corticotropin releasing factor binding protein (CRF-BP) is expressed in neuronal and non-neuronal cells. Brain Res., **698**, 259-264.
- 19. De Souza, E.B., Lovenberg, T.W., Chalmers, D.T., Grigoriadis, D.E., Liaw, C.W., **Behan, D.P.** and McCarthy, J. (1995). Heterogeneity of corticotropin releasing factor receptors: multiple targets for the treatment of CNS and inflammatory disorders. Annual Reports In Medicinal Chemistry, **30**, 21-30.
- 20. Sutton, S.W., **Behan, D.P.**, Larichi, S.L., Kaiser, R., Corrigan, A., Lowry, P.J., Potter, E., Perrin, M.H., Rivier, J. and Vale, W. (1995). Ligand requirements of the human corticotropin-releasing factor-binding protein. Endocrinology, **136**, 1097-1102.
- 21. **Behan, D.P.**, De Souza, E.B., Potter, E., Sawchenko, P., Lowry, P.J. and Vale, W.W. (1996). Modulatory actions of corticotropin releasing factor binding protein. Ann. New York Acad. Sci., 780, 81-95
- 22. **Behan, D.P.**, Grigoriadis, D.E., Lovenberg, T., Chalmers, D., Heinrichs, S., Liaw, C. and De Souza, E.B. (1996). Neurobiology of corticotropin releasing factor (CRF) receptors and CRF binding protein: implications for the treatment of CNS disorders. Molecular Psychiatry, 1, 265-277.
- 23. **Behan , D.P.**, Heinrichs, S.C. and De Souza, E.B. (1996). Corticotropin releasing factor (CRF) and Alzheimer's disease: recent developments with implications for more effective therapies. Expert Opinion In Investigational Drugs, **5**, 1277-1289.
- 24. **Behan, D.P.**, Khongsaly, O., Ling, N. and De Souza, E.B. (1996). Urocortin interaction with corticotropin releasing factor (CRF) binding protein (CRF-BP): a novel mechanism for elevating "free" CRF levels in human brain. Brain Res., 725, 263-267.
- 25. Chalmers, D.T., Lovenberg, T.W., Grigoriadis, D.E., **Behan, D.P.** and De Souza, E.B. (1996). Corticotropin-releasing factor receptors: from molecular cloning to drug design. Trends In Pharmacological Sciences, **17**, 166-172.
- 26. Heinrichs, S., Lapsansky, J., Behan, D.P, Chan, R.K., Sawchenko, P.E, Lorang, M.,

- Ling N., Vale, W.W. and De Souza, E. (1996). Corticotropin-releasing factor-binding protein ligand inhibitor blunts excessive weight gain in genetically obese Zucker rats and rats during nicotine withdrawal. Proc. Natl. Acad. Sci. (USA), 93, 15475-15480.
- 27. Heinrichs, S.C., Vale, E.A., Lapsansky, J., **Behan, D.P.**, McClure, L.V., Ling, N., De Souza, E.B. and Schulteis, G. (1996). Enhancement of performance in multiple learning tasks by corticotropin-releasing factor-binding protein ligand inhibitors. Peptides, **18**, 711-716.
- 28. Lowry, P.J., Koerber, S.C., Woods, R., Baigent, S., Sutton, S., **Behan, D.P.**, Vale, W. and Rivier, J. (1996). Nature of ligand affinity and dimerisation of corticotrophin-releasing factor binding protein may be detected by circular dichroism. J. Mol. Endocrinol., 16, 39-44.
- 29. Maciejeski, D., Crowe, P., De Souza, E.B. and **Behan, D.P.** (1996). Regulation of corticotropin releasing factor-binding protein (CRF-BP) expression in cultured rat astrocytes. J. Pharmacol. & Expt. Therap., **278**, 455-461.
- 30. **Behan, D.P.**, Khongsaly, O., Owens, M., Nemeroff, C.B. and De Souza, E.B. (1997). Measurement of corticotropin releasing factor (CRF), CRF-binding protein (CRF-BP) and CRF/CRF-BP complex in human brain: implications for Alzheimer's disease. J. Neurochemistry, **68**, 2053-2060.
- 31. Loddick, S.A., Liu, X-J., Lu, Z., Liu, C., **Behan, D.P.**, Chalmers, D.C., Foster, A.C., Vale, W.W., Ling, N., De Souza, E. (1998). Displacement of insulin-like growth factors from their binding proteins as a potential treatment for stroke. Proc. Natl. Acad. Sci. (USA), 95, 1894-1898.
- 32. Kasckow JW, Lupien SJ, **Behan DP**, Welge J, Hauger RJ.Circulating human corticotropin-releasing factor-binding protein levels following cortisol infusions. Life Sci. 2001 Jun 1;69(2):133-42
- 33. **Dominic P Behan** and Derek T Chalmers. The use of constitutively active receptors for drug discovery at the G protein-coupled receptor gene pool. Current Opinion in Drug Discovery & Development, 4(5):548-560, 2001.
- 34. Menzaghi, F., **Behan, D.P.** & Chalmers, D.T. Constitutively activated G protein-coupled receptors: a novel approach to CNS drug discovery. Curr. Drug Targets, **1**, 105-121, 2002.
- 35. Derek T. Chalmers and **Dominic P. Behan.** The Use of Constitutively Active GPCRs in Drug Discovery and Functional Genomics. Nature Reviews, 1, 599-608, 2002.
- 36. McLean, A.J., Zeng, F.Y., **Behan, D.**, Chalmers, D. and Milligan, G. (2002). Generation and analysis of constitutively active and physically destabilized mutants of the human beta(1)-adrenoceptor. Molecular Pharmacology, 62, 747-755.

- 37. Hakak, Y., Shrestha, D., Goegel, M.C., **Behan, D.P.**, Chalmers, D.T. (2003). Global analysis of G-protein-coupled receptor signaling in human tissues. FEBS Lett., 550, 11-17.
- 38. Zeng, F.Y., McLean, A.J., Milligan, G., Lerner, M., Chalmers, D.T., **Behan, D.P.** (2003) Ligand specific up-regulation of a Renilla reniformis luciferase-tagged, structurally unstable muscarinic M3 chimeric G protein coupled receptor. Molecular Pharmacology, 64, 1474-1484.

Abstracts & International Society Presentations

- 1. **Behan, D.P.**, Kennard, J.H. and Linton, E.A. (1987). Human plasma carrier-bound CRF-41 has biological activity. J. Endocrinol. Invest. 10 [suppl. 3], 63.
- 2. **Behan, D.P.**, Linton, E.A. and Lowry, P.J. (1987). Isolation of the human plasma CRF-binding protein by affinity chromatography. Physiology society meeting, London, December 1987.
- 3. **Behan, D.P.,** Linton, E.A. and Lowry, P.J. (1988). Isolation of the human plasma CRH-binding protein using affinity chromatography. XV111 Colloque Socie'te' de Neuroendocrinologie Experimentale. Annales d' Endocrinologie, Abstract No. 15.
- 4. **Behan, D.P.,** Linton, E.A. and Lowry, P.J. (1988). The human plasma CRF-carrier: Its isolation using affinity chromatography. 7th joint meeting of the British Endocrine Societies, Exeter University, England.
- 5. Lowry, P.J., **Behan, D.P.** and Linton, E.A. (1988). Central and peripheral control of neuropeptide gene expression. 8th International Congress of Endocrinology. July 1988, Kyoto, Japan, Abstract S-1.
- 6. **Behan, D.P.**, Linton, E.A. and Lowry, P.J. (1989). Central and peripheral regulation of neuropeptide gene expression. Proceedings of the 8th International Congress of Endocrinology. July 1988, Kyoto, Japan.
- 7. Potter, E, **Behan, D.P.**, Lowry, P. and Vale, W. (1993). HPA axis can alter the expression of corticotropin-releasing factor binding protein in the pituitary. In 75th Annual Meeting of the Endocrine Society. Las Vegas; Abstract 1675.
- 8. **Behan, D. P.,** Troncoso, J. C., Ling, N. and De Souza, E. B. (1994). Corticotropin releasing factor-binding protein in human brain: identification and characterization in Alzheimer's disease. Soc. Neurosci. Abstr. 20:552.10
- 9. Cepoi, D., Behan, D., Fischer, W.H., Lowry, P. and Vale, W.W. (1994). Cloning and characterization of corticotropin-releasing factor binding proteins form sheep brain. The Endocrine Society 15-18 June 1994.

- 10. De Souza, E. B., Grigoriadis, D. E. and **Behan, D. P.** (1994). Corticotropin releasing factor receptors and binding protein in the brain-endocrine-immune axis: role in CNS and immune disorders. 25th Congress of the International Society of Psychoneuroendocrinology, Seattle, WA, August 17-19.
- 11. Reynolds, P. J., Grigoriadis, D. E., Lovenberg, T. W., Wong, T. T., **Behan, D. P.**, De Souza, E. B. and Conlon, P. J. (1994). Presence of corticotropin-releasing factor (CRF) receptors on human peripheral blood cells and monocyte lines. Soc. Neurosci. Abstr. 20:552.9.
- 12. **Behan, D. P.**, Liu, X-J., Ling, N., Maciejewski, D., Chalmers, D., Nasman, B., Olsson, T., Troncoso, J. and De Souza, E. B. (1995). Corticotropin releasing factor (CRF) and CRF-binding protein in Alzheimer's disease. Summer Neuropeptide Conf. Abstr., Martha's Vineyard, June 24 -27.
- 13. **Behan, D. P.**, Liu, X-J., Ling, N., Nasman, B., Olsson, T., Troncoso, J. and De Souza, E. B. (1995). Direct measurement of corticotropin releasing factor-binding protein and CRF-BP/CRF complex in Alzheimer's disease using two-site ELISA assays. Soc. Neurosci. Abstr. 21:532.5.
- 14. De Souza, E. B., **Behan, D. P.** and Heinrichs, S. P. (1995). Corticotropin releasing factor (CRF)-binding protein (CRF-BP): a novel target for the symptomatic treatment of cognitive deficits in Alzheimer's disease. American College of Neuropsychopharmacology, San Juan, Puerto Rico, p. 29.
- 15. Heinrichs, S. C., **Behan, D. P.**, Ling, N. and De Souza, E. B. (1995). Corticotropinreleasing factor binding protein ligand inhibition: A novel mechanism for cognitive enhancement. Soc. Neurosci. Abstr. 21: 532.7.
- 16. Maciejewski, D. L., **Behan, D.**, Chalmers, D. T. and De Souza, E. B. CRF binding protein (CRF-BP) is expressed in cultured astrocytes and neurons in response to cAMP and protein kinase C stimulation. (1995). Soc. Neurosci. Abstr. 21:532.6.
- 17. **Behan, D.P.**, Heinrichs, S. and De Souza, E.B. Interactions between corticotropin releasing factor (CRF) and CRF-binding protein (CRF-BP) in Alzheimer's disease. (1996). British Neuroendocrine Group. Annual Meeting. 2nd-3rd July, Edinburgh (invited symposium speaker).
- 18. De Souza, E.B., Lapsansky, J., **Behan, D.P.**, Chan, R.K.W., Sawchenko, P.E., Lorang, M., Ling, N., Vale, W.W. and Heinrichs, S.C. Corticotropin releasing factor-binding protein ligand inhibitors as a potential treatment for excessive weight gain associated with obesity and smoking cessation. ACNP, 1996.
- 19. Heinrichs, S., Lapsansky, J., **Behan, D.**, Chan, R., Sawchenko, P., Lorang, M., Ling, N., Vale, W., and De Souza, E. Role of CRF in Energy Balance. New Drug Targets for

the Treatment of Obesity, Cambridge, MA, December 1996.

4)

- 20. Heinrichs, S., Lapsansky, J., **Behan, D.,** Chan, R., Sawchenko, P., Lorang, M., Ling, N., Vale, W. and De Souza, E. Role of CRF in Energy Balance. Obesity: Advances in Therapeutics and Drug Development, San Diego, CA, January 1997.
- 21. Lapsansky, J.S., Vale, E.A., **Behan, D.P.**, McClure, L.V., Ling, N., De Souza, E.B., Schulteis, G. and Heinrichs, S.C. Enhancement of performance in multiple learning tasks by corticotropin releasing factor-binding protein ligand inhibitors. Annual meeting of the Society for Neuroscience, New Orleans, 1997.
- 22. Liaw, C., Bruinsma, K., Thomsen, B., Russo, J., **Behan, D.**, Chalmers, D., Herrick-Davis, K., Egan, C., Grinde, E. and Teitler, M. Antipsychotic agents possessing inverse agonist activity at the human 5-HT_{2C} receptor. Annual meeting of the Society of Neuroscience, Los Angeles, 1998.
- 23. Thomsen, W.J., Russo, J.F., Bruinsma, K., Liaw, C.W., Glen, R.C., Smith, J.R., Chalmers, D.T., and **Behan, D.P.** Use of constitutively activated human 5HT_{2C} and 5HT_{2A} receptors to directly identify inverse agonists: evaluation of standard reference compounds. 29th meeting of The Society for Neuroscience, Miami Beach, Florida, October 23-28, 1999, Abstract # 483.3.
- 24. Russo, J.F., Thomsen, W.J., Reyes, H.S., Ravelle, G.P., Bruinsma, K., Liaw, C.W., Glen, R.C., Smith, J.R., Chalmers, D.T. and **Behan D.P**. Direct identification of high affinity inverse agonists using constitutively active 5HT_{2C} and 5HT_{2A} receptors. 29th meeting of The Society for Neuroscience, Miami Beach, Florida, October 23-28, 1999, Abstract # 483.4.
- 25. Warmoth, K., Trujillo, K.A., Ruzek, E., Glen, R., Smith, J., Thomsen, W., **Behan, D.P.** and Chalmers, D.T. AR116,102, A novel high affinity 5HT2A receptor inverse agonist with in vivo efficacy. 29th meeting of The Society for Neuroscience, Miami Beach, Florida, October 23rd-28th, 1999, Abstract # 484.4.
- 26. Hittner, J.M., Whelan, K.T., Trujillo, K.A., Thomsen, W.J., Russo, J.F., Reyes, H.S., Glen, R.C., Smith, J.R., Beeley, N.P., **Behan, D.P.**, Chalmers, D.T. and Menzaghi, F. A selective 5-HT2A receptor inverse agonist with preclinical antipsychotic profile in rats. The Society for Neuroscience, New Orleans, Lousisiana, Nov 4th-9th, 2000. Abstract 534.12.
- 27. Menzaghi, F., Whelan, K.T., Trujillo, K.A., Hittner, J., Thomsen, W.J., Beeley, N., Hodgkin, E.E., Glen, R.C., Smith, J.R., **Behan, D.P.** and Chalmers, D.T.. AR116081, A novel selective 5-HT2A inverse agonist as a putative atypical antipsychotic: comparative studies with clozapine and haloperidol. CINP 2000.
- 28. Menzaghi, F., Trujillo, K.A., Thomsen, W.J., Whelan, K.T., Russo, J.F., Reyes, H.S., Beeley, N., Liaw, C.W., Hodgkin, E.E., Glen, R.C., Smith, J.R., Behan, D.P. and

- Chalmers, D.T. Identification of novel novel selective 5-HT2A inverse agonists as putative atypical antipsychotics using constitutively activated human 5-HT receptors. American Society for Pharmacology and Experimental Therapeutics (ASPET) 2000.
- 29. Menzaghi, F., Whelan, K.T., Hittner, J.M., McDonald, J.S., Risbrough, V.B., Trujillo, K.A., Lu, X.Y., Villegas, S., Gore, M., Liaw, C.W., Beeley, N.P., Watson, S.J., **Behan, D.P.** and Chalmers, D.T. The orphan G-protein-coupled receptor ARE113 is a novel receptor target for obesity. The Society for Neuroscience, New Orleans, Lousiana, Nov 4th-9th, 2000. Abstract 569.18.
- 30. Ortuno, D., Xu, S., White, C., Liaw, C., **Behan, D.**, Chalmers, D., Gore, M. The orphan GPCRs 18ARK and 19AM are expressed by myelinating cells of the PNS and CNS. The 30th annual meeting for the Society of Neuroscience, November 4th-9th, 2000, New Orleans. Abstract 517.2.
- 31. Rachmeler, H., Gore, M., **Behan, D.**, Chalmers, D. The chemokine receptor related orphan GPCR, CRRO1, is induced in microglial cells following LPS treatment. The 30th annual meeting for the Society of Neuroscience, November 4th-9th, 2000, New Orleans. Abstract 606.26.
- 32. Chen, R., Dang, H., Leonard, J., Chalmers, D., **Behan, D.** Identification and characterization of a second melanin-concentrating hormone receptor. 31st annual meeting of the Society for Neuroscience, 10th-15th November, San Diego, 2001. Abstract 461.6.
- 33. Hakak, Y., Leonard, J., Bagnol, D., **Behan**, **D**., Chalmers, D.T.. Genome-wide analysis of G-protein coupled receptors in the human brain using a GPCR oligonucleotide microarray. 31st annual meeting of the Society for Neuroscience, 10th-15th November, San Diego, 2001. Abstract 264.3.
- 34. Maciejewski-Lenoir, D., Guerra, N.C., **Behan**, **D**., Chalmers, D., Gore, M. 19 AA, an orphan GPCR closely related to the n-formyl peptide receptor family, is expressed in rat cultured microglia upon INF stimulation. 31st annual meeting of the Society for Neuroscience, 10th-15th November, San Diego, 2001. Abstract 899.15.
- 35. Ortuno, D., White, C.A., Liaw, C., **Behan, D.**, Chalmers, D.T., Gore, M. Expression of GPCRs in myelinating glia in the CNS and the PNS may provide therapeutic targets for central and peripheral neuropathies. 31st annual meeting of the Society for Neuroscience, 10th-15th November, San Diego, 2001. Abstract 103.17.
- 36. Bagnol, D., Hakak, Y., Ortuno, D., Zhou, V., Shrestha, D., **Behan, D**. and Chalmers, D. Global analysis of GPCRs in human hypothalamic neurons involved in the regulation of food intake using a GPCR oligonucleotide microarray. Federation of European Neurosciences (FENS), 2002.

- 37. Hakak, Y., Bagnol, D., Shrestha, D., Chu, M.F., **Behan, D.** and Chalmers, D. Microarray analysis of GPCR expression in human midbrain dopamine neurons isolated by laser capture microscopy. 32nd annual meeting of the Society for Neuroscience, 2nd-7th November, Orlando, Florida, 2002.
- 38. Maciejewski-Lenoir, D., Shrestha, D., Bagnol, D., Zhou, V.W., **Behan, D.**, Chalmers, D. and Hakak, Y. Microarray analysis of GPCR expression in human stem/progenitor cell populations 32nd annual meeting of the Society for Neuroscience, 2nd-7th November, Orlando, Florida, 2002.
- 39. Menzaghi F, Whelan KT, Thomsen WJ, Beeley N, Glen R, **Behan DP** and Chalmers DT. Therapeutic potential of selective serotonin 5HT2A receptor inverse agonists: Pre-clinical evaluation of AR116081 as antipsychotic in rodents. Federation of European Neurosciences (FENS), 2002.

Recent meetings and presentations

International Business Communications IBC's Target Validation Conference: Invited speaker. April 23-25, San Diego Paradise Point Resort, San Diego, CA.

AstaZeneca sponsored Peptide Receptor Conference. Invited Speaker. Montreal, July 21, 2001.

The Center for Business Intelligence (CBI). Optimizing Lead Selection and Early Attrition. Invited Speaker and Chairman. July 23, 2001 Adam's Mark Philadelphia.

International Business Communications (IBCs) 6th Annual World Congress. Invited speaker. August The GPCR Factory. 12-17, 2001, Boston, MA. The World Trade Center and Seaport Hotel.

Patents

Named inventor on over 100 patents/patent applications, including:

- Potter, E., **Behan, D.P.**, Fischer, W.H., Linton, E.A., Lowry P.J., Vale, W.W. DNA Encoding CRF Binding Protein. U.S. Patent No. 5,464,757.
- Behan, D.P., Vale, W.W., Fischer, W.H., Lowry, P.J. Brain-Derived Membrane-Associated CRF Binding Proteins. U.S. Patent No. 5,587,462.
- Potter, E., **Behan, D.P.**, Linton, E.A., Lowry, P.J., Vale, W.W. CRF Binding Protein Antibodies and Assays Using Same. U.S. Patent No. 5,733,790.
- Potter, E., **Behan, D.P.**, Fischer, W.H., Linton, E.A., Lowry, P.J., Vale, W.W. CRF Binding Protein. U.S. Patent No. 5,844,080.
- Behan, D.P., Vale, W.W., Fischer, W.H., Lowry, P.J. Brain-Derived Membrane-Associated CRF Binding Proteins. U.S. Patent No. 5,910,428.

- Whitten, J.P., McCarthy J.R., Liu, Z., Huang, C.Q., Erickson, P.E., **Behan, D.P.** Compounds and Methods for Increasing Endogenous Levels of Corticotropin-Releasing Factor. U.S. Patent No. 5,959,109.
- Behan, D.P., Chalmers, D.T., Foster, R.J., Glen, R.C., Lawless, M.S., Liaw, C.W., Liu, Q., Russo, J.F., Smith, J.R., Thomsen, W.J. 5-HT2A Receptor Inverse Agonists. U.S. Patent No. 6,107,324.
- Whitten, J.P., McCarthy, J.R., Liu, Z., Huang, C.Q., Erickson, P.E., **Behan, D.P.** Compounds and Methods for Increasing Endogenous Levels of Corticotropin-Releasing Factor. U.S. Patent No. 6,133,276.
- Behan, D.P., Chalmers, D.T., Foster, R.J., Glen, R.C., Lawless, M.S., Liaw, C.W., Liu, Q., Russo, J.F., Smith, J.R., Thomsen, W.J. Non-Endogenous, Constitutively Activated Human Serotonin Receptors and Small Molecule Modulators Thereof. U.S. Patent No. 6,140,509.
- **Behan, D.P.**, Chalmers, D.T., Beeley, N.R.A., Foster, R.J., Glen, R.C., Lawless, M.S., Liaw, C.W., Liu, Q., Menzaghi, F., Russo, J.F., Smith, J.R., Thomsen, W.J. Small Molecule Modulators of Non-Endogenous, Constitutively Activated Human Serotonin Receptors. U.S. Patent No. 6,150,393.
- **Behan, D.P.**, Chalmers, D.T., Liaw, C.W., Russo, J.F., Thomsen, W.J. Non-Endogenous, Constitutively Activated Human Serotonin Receptors and Small Molecule Modulators Thereof. U.S. Patent No. 6,420,541.
- Beeley, N.R.A., **Behan, D.P.**, Chalmers, D.T., Menzaghi, F., Strah-Pleynet, S. Small Molecule Modulators of G Protein-Coupled Receptor Six. U.S. Patent No. 6,420,563.
- **Behan, D.P.**, Chalmers, D.T., Liaw, C.W., Russo, J.F., Thomsen, W.J. Non-Endogenous, Constitutively Activated Human Serotonin Receptors and Small Molecule Modulators Thereof. U.S. Patent No. 6,541,209.
- Liaw, C.W., **Behan, D.P.**, Chalmers, D.T. Non-Endogenous, Constitutively Activated Human G Protein-Coupled Receptors. U.S. Patent No. 6,555,339.
- Behan, D.P., Chalmers, D.T., Liaw, C.W., Lin-Lin, I., Lowitz, K.P., Chen, R. Endogenous Constitutively Activated G Protein-Coupled Orphan Receptors. U.S. Patent No. 6,653,086.

and secretary pathways that have evolved for mammalian systems – thus, results obtained in non-mammalian cells, while of potential use, are not as preferred as that obtained from mammalian cells. Of the mammalian cells, COS-7, 293 and 293T cells are particularly preferred, although the specific mammalian cell utilized can be predicated upon the particular needs of the artisan.

On day one, 1X10⁷ 293T cells per 150mm plate were plated out. On day two, two reaction tubes were prepared (the proportions to follow for each tube are per plate): tube A was prepared by mixing 20µg DNA (e.g., pCMV vector; pCMV vector with receptor cDNA, etc.) in 1.2ml serum free DMEM (Irvine Scientific, Irvine, CA); tube B was prepared by mixing 120µl lipofectamine (Gibco BRL) in 1.2ml serum free DMEM. Tubes A and B were then admixed by inversions (several times), followed by incubation at room temperature for 30-45min. The admixture is referred to as the "transfection mixture". Plated 293T cells were washed with 1XPBS, followed by addition of 10ml serum free DMEM. 2.4ml of the transfection mixture was then added to the cells, followed by incubation for 4hrs at 37°C/5% CO₂. The transfection mixture was then removed by aspiration, followed by the addition of 25ml of DMEM/10% Fetal Bovine Serum. Cells were then incubated at 37°C/5% CO₂. After 72hr incubation, cells were then harvested and utilized for analysis.

Example 4 REPORTER-BASED ASSAY: CRE-LUC REPORTER ASSAY

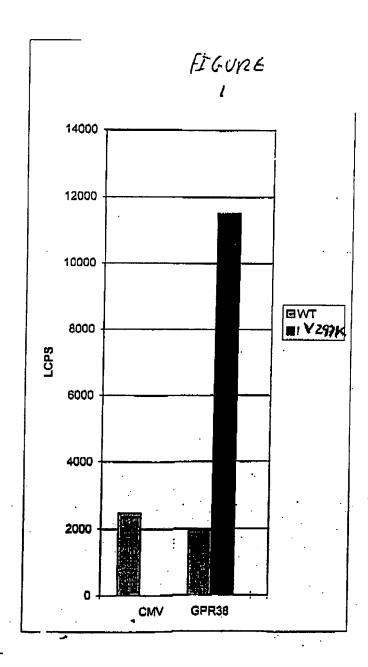
A method to detect Gs stimulation depends on the known property of the transcription factor CREB, which is activated in a cAMP-dependent manner. A PathDetect pCRE-Luc trans-Reporting System (Stratagene, Catalogue # 219075) was utilized to assay for Gs coupled activity in 293T cells. Cells were transfected with the

plasmids components of this system and the indicated expression plasmid encoding endogenous or non-endogenous receptor using a Mammalian Transfection Kit (Stratagene, Catalogue #200285) according to the manufacturer's instructions. Briefly, 400 ng pCRE-Luc, 80 ng pCMV (comprising the receptor) and 20 ng CMV-SEAP (secreted alkaline phosphatase expression plasmid; alkaline phosphatase activity is measured in the media of transfected cells to control for variations in transfection efficiency between samples) were combined in a calcium phosphate precipitate as per the manufacturer's instructions. Half of the precipitate was equally distributed over 3 wells in a 96-well plate, kept on the cells overnight, and replaced with fresh medium the following day. Forty-eight (48) hr after the start of the transfection, cells were treated and assayed for luciferase activity using a LucliteTM Kit (Packard, Cat. # 6016911) and "Trilux 1450 Microbeta" liquid scintillation and luminescence counter (Wallac) as per the manufacturer's instructions. The data was analyzed using GraphPad PrismTM 2.0a (GraphPad Software Inc.). Results are summarized in Figure 1.

Figure 1 represents an 83.1% increase in activity of the non-endogenous, constitutively active version of human GPR38 (V297K) (11,505 relative light units) compared with that of the endogenous GPR38 (1950 relative light units).

References cited throughout this patent document, unless otherwise indicated, are incorporated herein by reference. Modifications and extension of the disclosed inventions that are within the purview of the skilled artisan are encompassed within the above disclosure and the claims that follow.

Although a variety of expression vectors are available to those in the art, for purposes of utilization for both the endogenous and non-endogenous human GPR38, it is



GOODMAN & GILMAN'S The PHARMACOLOGICAL BASIS OF THERAPEUTICS

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THYROID AND ANTITHYROID DRUGS

Alan P. Farwell and Lewis E. Braverman

This chapter discusses the function of the thyroid hormones, thyroxine (T_4) and triiodothyronine (T_3) , in growth and metabolism and the regulation of thyroid function by
thyroid-stimulating hormone (TSH) secreted from the pituitary. Calcitonin, also secreted
by the thyroid gland, is discussed in Chapter 61. Evaluation of free thyroxine and TSH
levels as a means to assess thyroid function is provided as a prelude to the discussion of
treatment of the hypothyroid patient with hormone replacement and of the hyperthyroid
individual with one of a variety of antithyroid drugs, such as propylthiouracil and methimazole, and other thyroid inhibitors, including ionic inhibitors that interfere with the concentration of iodide by the thyroid gland and radioactive iodine, used both for diagnosis
as well as treatment of hypothyroidism. Although disorders of the thyroid are common,
effective treatment of most thyroid disorders is available.

Thyroid hormones, the only known iodine-containing compounds with biological activity, have two important functions. In developing animals and human beings, they are crucial determinants of normal development, especially in the central nervous system (CNS). In the adult, thyroid hormones act to maintain metabolic homeostasis, affecting the function of virtually all organ systems. To meet these requirements, there are large stores of preformed hormone within the thyroid gland. Metabolism of the thyroid hormones occurs primarily in the liver, although local metabolism within certain target tissues, such as the brain, also occurs. Serum concentrations of thyroid hormones are precisely regulated by the pituitary hormone, thyrotropin, in a classic negative-feedback system. The predominant actions of thyroid hormone are mediated via binding to nuclear thyroid hormone receptors and modulating transcription of specific genes. In this regard, thyroid hormones share a common mechanism of action with steroid hormones, vitamin D, and retinoids, whose receptors make up a superfamily of nuclear receptors (see Chapter 2).

Disorders of the thyroid are common. They consist of two general presentations: changes in the size or shape of the gland or changes in secretion of hormones from the gland. Thyroid nodules and goiter in the euthyroid patient are the most common endocrinopathies and can be caused by benign and malignant tumors. The presentation of overt hyper- or hypothyroidism often presents the clinician with dramatic clinical manifestations. While the diagnosis may

be clinically obvious, subtle presentations require the use of biochemical tests of thyroid function. Screening of the newborn population for congenital hypothyroidism, followed by the institution of appropriate thyroid hormone replacement therapy, has dramatically decreased the incidence of mental retardation and cretinism in the United States. Worldwide, congenital hypothyroidism due to iodine deficiency remains the major preventable cause of mental retardation.

Effective treatment of most thyroid disorders is readily available. Treatment of the hypothyroid patient is straightforward and consists of hormone replacement. There are more options for treatment of the hyperthyroid patient, including the use of antithyroid drugs to decrease hormone synthesis and secretion by the gland and destruction of the gland by the administration of radioactive iodine or by surgical removal. Treatment of thyroid disorders in general is extremely satisfying, as most patients can be either cured or have their diseases controlled (see Braverman and Utiger, 1991; Braverman and Refetoff, 1994).

THYROID

The thyroid gland is the source of two fundamentally different types of hormones. The iodothyronine hormones include thyroxine and 3,5,3'-triiodothyronine; they are essential for normal growth and development and play an important role in energy metabolism. The other known secretory product of the thyroid, calcitonin, is produced by the parafollicular (C-) cells and is discussed in Chapter 61.

History. The thyroid gland was first described by Galen and was named "glandulae thyroidaeae" by Wharton in 1656. Harington (1935) reviewed the many older opinions concerning the function of this gland. Wharton thought, for example, that the viscous fluid within the follicles lubricated the trachea. He also believed that the gland was larger in women to serve a cosmetic function in giving grace to the contour of the neck. Later observers, influenced by the liberal blood supply of the gland, believed that it provided a vascular shunt for the brain. With this function in mind, Rush in 1820 expressed the belief that the larger size of the gland in women was "necessary to guard the female system from the influence of the more numerous causes of irritation and vexation of mind to which they are exposed than the male sex." However, Hofrichter opposed this theory in the same year by pointing out that "If it were indeed true that the thyroid contains more blood at some times than at others, this effect would be visible to the naked eye; in this case women would certainly have long ceased to go about with bare necks, for husbands would have learned to recognize the swelling of this gland as a danger signal of threatening trouble from their better halves."

The thyroid was first recognized as an organ of importance when enlargement was observed to be associated with changes in the eyes and the heart in the condition we now call hyperthyroidism. It is of interest that this condition, the manifestations of which on occasion an be as striking as any in medicine, escaped description until Parry saw his first case in 1786. Parry's account was not published until 1825 and was followed in 1835 and 1840 by those of Graves and Basedow, whose names became applied to the disorder. In 1874 Gull first associated atrophy of the gland with the symptoms now known to be characteristic of thyroid deficiency, and hypofunction of the thyroid, hypothyroidism, in adults was known as Gull's disease. The term myxedema was applied to the clinical syndrome in 1878 by Ord in the belief that the characteristic thickening of the subcutaneous tissues was due to excessive formation of mucus.

Extirpation experiments to elucidate the function of the thyroid were at first misinterpreted because of the simultaneous removal of the parathyroids. However, the pioneer research in the late 19th century on the latter organs by Gley allowed the functional differentiation of these two endocrine glands. It was not until after calcitonin was discovered in 1961 that it was realized that the thyroid itself also was concerned with the regulation of Ca²⁺. In 1891, Murray became the first to treat a case of hypothyroidism by injecting an extract of the thyroid gland; in the following year, Howitz, Mackenzie, and Fox independently discovered that thyroid tissue was fully effective when given by mouth.

Magnus-Levy discovered the effect of the thyroid on metabolic rate in 1895; he found that Gull's disease was characterized by a low rate of metabolism and that the administration of thyroid to hypothyroid or normal individuals increased oxygen consumption.

Chemistry of Thyroid Hormones. The principal hormones of the thyroid gland are the iodine-containing amino acid derivatives of thyronine—thyroxine (T₄) and T₃ (triiodothyronine; 3,5,3'-triiodothyronine; Figure 56-1). Thyroxine was first isolated in crystalline form om a hydrolysate of thyroid by Kendall in 1915; he found that the crystalline product exerted the same physiological effects as the extract from which it was obtained. Eleven years later the structural

Thyronine CH2CHCOOH Thyroxine NH2 3,5,3'-Triiodothyronine CH2CHCOOH NH₂ 3,3',5'-Triiodothyronine CH2 CHCOOH NH₂ Diiodotyrosine 12 CHCOOH lodotyrosine H2CHCOOH NH₂

th

Figure 56-1. Thyronine, thyroid hormones, and precursors.

formula of thyroxine was elucidated by Harington, and in 1927 Harington and Barger synthesized the hormone.

Following the isolation and the chemical identification of thyroxine, it was generally believed that all the hormonal activity of thyroid tissue could be accounted for by its content of thyroxine. However, careful studies revealed that crude thyroid preparations possessed greater calorigenic activity than could be accounted for by their thyroxine content. The enigma was resolved with the detection, isolation, and synthesis of triiodothyronine (Gross and Pitt-Rivers, 1952; Roche et al., 1952a, 1952b). Further studies revealed that triiodothyronine is qualitatively similar to thyroxine in its biological action but that it is much more potent on a molar basis (Gross and Pitt-Rivers, 1953a, 1953b).

Structure-Activity Relationship. The stereochemical nature of the thyroid hormones plays an important role in defining hormone activity. A great many structural analogs of thyroxine have been synthesized in order to define the structure-activity relationship, to detect antagonists of thyroid hormones, or to find compounds exhibiting one desirable type of activity while not showing unwanted effects.

The only significant success has been the partial separation of the cholesterol-lowering action of thyroxine analogs from their calorigenic or cardiac effects. For example, introduction of specific arylmethyl groups at the 3' position of triiodothyronine resulted in analogs that are liver-selective, cardiac-sparing thyromimetics (Leeson et al., 1989). The D isomer of thyroxine was once used to lower the concentration of cholesterol in plasma, but cardiac side effects resulted in discontinuation of the clinical uses of this hormone. Newer analogs offer hope that more useful separation of these activities may yet be achievable (Underwood et al., 1986; Sherman and Ladenson, 1992).

The structural requirements for a significant degree of thyroid hormone activity have been defined (see Jorgensen, 1964; Cody, 1980, 1991). The 3'-monosubstituted compounds are more active than the 3',5'-disubstituted molecules. Thus, triiodothyronine is five times more potent than thyroxine, while 3'-isopropyl-3,5-diiodothyronine has seven times the activity.

Although the chemical nature of the 3, 5, 3', and 5' substituents is important, their effects on the conformation of the molecule are even more so. In thyronine, the two rings are angulated at about 120° at the ether oxygen and are free to rotate on their axes. As depicted schematically in Figure 56–2, when the 3,5 iodines are in place, rotation of the two rings is somewhat restricted, and they tend to take up positions perpendicular to one another. While not potent, even halogen-free derivatives possess some activity if they have the proper conformation. In general, the affinity of iodothyronines for the thyroid hormone receptor parallels their biological potency (Oppenheimer et al., 1987), but additional factors including affinity for plasma proteins, rate of entry into cell nuclei, and rate of metabolism can affect therapeutic potency.

Recent structure-activity correlations indicate that certain plant flavonoids that are long-standing folk remedies can exhibit antihormonal properties, including inhibition of the enzyme that catalyzes 5' (outer, or tyrosyl ring) deiodination of T_4 (type I iodothyronine 5'-deiodinase; Cody, 1991). These compounds are also potent competitors of thyroxine binding to transthyretin. Computer graphic modeling suggests that the best structural homology between thyroid hormones and flavonoids involves their respective phenolic rings.

Synthesis of Thyroid Hormones.—The synthesis of the thyroid hormones is unique, complex, and seemingly grossly inefficient. The thyroid hormones are synthesized and stored as amino acid residues of thyroglobulin, a pro-

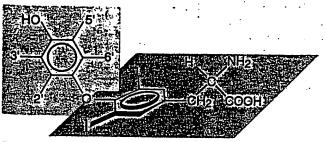


Figure 56-2. Structural formula of 3,5-diiodothyronine, drawn to show the conformation in which the planes of the aromatic rings are perpendicular to each other. (Adapted from Jorgensen, 1964. See also Cody, 1980.)

tein constituting the vast majority of the thyroid follicular colloid. The thyroid gland is unique in storing great quantities of potential hormone in this way, and extracellular thyroglobulin can represent a large portion of the mass of the gland. Thyroglobulin is a complex glycoprotein made up of two apparently identical subunits, each with a molecular mass of 330 kDa. Interestingly, molecular cloning has revealed that thyroglobulin belongs to a superfamily of serine hydrolases, including acetylcholinesterase (see Chapter 8).

The major steps in the synthesis, storage, release, and interconversion of thyroid hormones are the following: (1) the uptake of iodide ion by the gland, (2) the oxidation of iodide and the iodination of tyrosyl groups of thyroglobulin, (3) coupling of iodotyrosine residues by ether linkage to generate the iodothyronines, (4) the proteolysis of thyroglobulin and the release of thyroxine and triiodothyronine into the blood, and (5) the conversion of thyroxine to triiodothyronine in peripheral tissues. These processes are summarized in Figure 56–3.

I. Uptake of Iodide. Iodine ingested in the diet reaches the circulation in the form of iodide. Under normal circumstances, its concentration in the blood is very low (0.2 to 0.4 µg/dl; about 15 to 30 nM), but the thyroid efficiently and actively transports the ion. As a result, the ratio of thyroid to plasma iodide concentration is usually between 20 and 50 and can far exceed 100 when the gland is stimulated. The iodide transport mechanism is inhibited by a number of ions such as thiocyanate and perchlorate (Figure 56-3). The transport system is stimulated by thyrotropin [thyroid-stimulating hormone (TSH); see below] and also is controlled by an autoregulatory mechanism. Thus, decreased stores of thyroid iodine enhance iodide uptake, and the administration-of-iodide-can reverse this situation.

If the further metabolism of iodide is blocked by antithyroid drugs, the iodide-concentrating mechanism can be more easily studied. Thus isolated, the mechanism resembles those found in other structures that concentrate iodide, including the salivary glands, gastric mucosa, midportion of the small intestine, choroid plexus, skin, mammary gland, and perhaps the placenta, all of which maintain a concentration of iodide greater than that of the blood. It has been suggested that the accumulation of iodide by the placenta and the mammary gland may be of importance in providing adequate supplies for the fetus and infant, but no obvious purpose is served by the accumulation of iodide at the other sites. It is evident that the iodide-accumulating system of the thyroid is not unique to the gland and does not account for the specific function of synthesizing thyroid hormone.

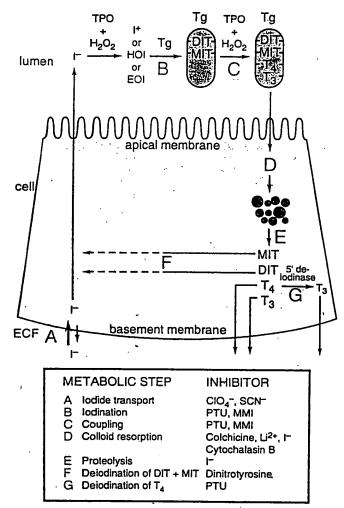


Figure 56-3. Major pathways of thyroid hormone biosynthesis and release.

Abbreviations are as follows: Tg, thyroglobulin; DIT, diiodotyrosine; MIT, monoiodotyrosine; TPO, thyroid peroxidase; HOI, hypoiodous acid; EOI, enzyme-linked species; PTU, propylthiouracil; MMI, methimazole; ECF, extracellular fluid. (Adapted from Taurog, 1991, with permission.)

2. Oxidation and Iodination. Consistent with the conditions generally necessary for halogenation of aromatic rings, the iodination of tyrosine residues requires the iodinating species to be in a higher state of oxidation than is the anion. The exact nature of the iodinating species was uncertain for many years. However, Magnusson and coworkers (1984) have provided convincing evidence that it is hypoiodate, either as hypoiodous acid (HOI) or as an enzyme-linked species (E-OI).

The oxidation of iodide to its active form is accomplished by thyroid peroxidase, a heme-containing enzyme that utilizes hydrogen peroxide (H₂O₂) as the oxidant (Tau-

rog, 1991; Magnusson et al., 1987). Thyroid peroxidase has been cloned and identified as an autoantigen in autoimmune thyroid disease (McLachlan and Rapoport, 1992). The peroxidase is membrane-bound and appears to be concentrated at or near the apical surface of the thyroid cell. The reaction results in the formation of monoiodotyrosyl and diiodotyrosyl residues in thyroglobulin just prior to its storage in the lumen of the thyroid follicle. It is thought that the formation of the H₂O₂ that serves as a substrate for the peroxidase occurs in close proximity to its site of utilization and involves the oxidation of reduced nicotinamide adenine di-nucleotide phosphate (NADPH). An increase in the generation of H₂O₂ may be an important facet of the mechanism by which TSH stimulates the organification of iodide in thyroid cells. This hypothesis has arisen from observations that TSH stimulates the synthesis of inositol trisphosphate and elevates cytosolic concentrations of Ca2+ in thyroid follicular cells (Corda et al., 1985; Field et al., 1987; Laurent et al., 1987); the formation of H₂O₂ is stimulated by a rise in cytosolic Ca²⁺ (Takasu et al., 1987).

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3. Formation of Thyroxine and Triiodothyronine from Iodotyrosines. The remaining synthetic step is the coupling of two diiodotyrosyl residues to form thyroxine or of monoiodotyrosyl and diiodotyrosyl residues to form triiodothyronine. These are also oxidative reactions and appear to be catalyzed by the same peroxidase discussed above. The mechanism involves the enzymatic transfer of groups, perhaps as iodotyrosyl free radicals or positively charged ions, within thyroglobulin. Although many other proteins can serve as substrates for the peroxidase, none is as efficient as thyroglobulin in yielding thyroxine. The configuration of the protein is thus presumed to be important in facilitating this coupling reaction. Thyroxine formation occurs primarily at a location near the amino terminus of the protein, while most of the triiodotyrosine is synthesized near the carboxy terminus (Dunn et al., 1987). The relative rates of synthetic activity at the various sites depend on the concentration of TSH and the availability of iodide. This may account, at least in part, for the long-known relationship between the proportion of thyroxine and triiodothyronine formed in the thyroid and the availability of iodide or the relative quantities of the two iodotyrosines. For example, when there is a deficiency of iodine in rat thyroid, the ratio of thyroxine to triiodothyronine decreases from 4:1 to 1:3 (Greer et al., 1968). Because triiodothyronine is at least five times as active as thyroxine and contains only/ three-fourths as much iodine, a decrease in the quantity of available iodine need have little impact on the effective amount of thyroid hormone elaborated by the gland. Although a decrease in the availability of iodide and

the associated increase in the proportion of monoiodotyrosine favor the formation of triiodothyronine over thyroxine, a deficiency in diiodotyrosine ultimately can impair the formation of both forms of the hormone. In addition to the coupling reaction, intrathyroidal and secreted triiodothyronine is generated by the 5'-deiodination of thyroxine (Chanoine et al., 1993).

4. Secretion of Thyroid Hormones. Since thyroxine and triiodothyronine are synthesized and stored within thyroglobulin, proteolysis is an important part of the secretory process. This process is initiated by endocytosis of colloid from the follicular lumen at the apical surface of the cell. This "ingested" thyroglobulin appears as intracellular colloid droplets, which apparently then fuse with lysosomes containing the requisite proteolytic enzymes. It is generally believed that thyroglobulin must be completely broken down into its constituent amino acids for the hormones to be released. As the molecular mass of thyroglobulin is 660 kDa, and the protein is made up of about 300 carbohydrate residues and 5500 amino acid residues, only two to five of which are thyroxine, this is an extravagant process. TSH appears to enhance the degradation of thyroglobulin by increasing the activity of several thiol endopeptidases of the lysosomes (Dunn and Dunn, 1988). The endopeptidases selectively cleave thyroglobulin, yielding hormone-containing intermediates that are subsequently processed by exopeptidases (Dunn el al., 1991). The liberated hormones then exit the cell, presumably at its basal membrane. When thyroglobulin is hydrolyzed, monoiodotyrosine and diiodotyrosine also are liberated, but they usually do not leave the thyroid. Instead, they are selectively metabolized, and the iodine, liberated in the form of iodide, is reincorporated into protein. Normally, all this iodide is reused; however, when proteolysis is activated intensely by TSH, some of the iodide reaches the circulation, at times accompanied by trace amounts of the iodotyrosines.

5. Conversion of Thyroxine to Triiodothyronine in Peripheral Tissues. The normal daily production of thyroxine has been estimated to range between 70 and 90 μ g, while that of triiodothyronine is between 15 and 30 μ g. Although triiodothyronine is secreted by the thyroid, metabolism of thyroxine by sequential monodeiodination in the peripheral tissues accounts for about 80% of circulating triiodothyronine (Figure 56-4). Removal of the 5'-, or outer ring, iodine leads to the formation of triiodothyronine and is the "activating" metabolic pathway. The major site of conversion of thyroxine to triiodothyronine outside the thyroid is the liver. Thus, when thyroxine is given to hypothyroid patients in doses that pro-

duce normal concentrations of thyroxine in plasma, the plasma concentration of triiodothyronine also reaches the normal range. Most peripheral target tissues utilize triiodothyronine that is derived from the circulating hormone. Notable exceptions are the brain and pituitary, for which local generation of triiodothyronine is a major source for the intracellular hormone. Removal of the iodine on position 5 of the inner ring produces the metabolically inactive 3,3',5'-triiodothyronine (reverse T₃, rT₃; Figure 56-1). Under normal conditions, about 41% of thyroxine is converted to triiodothyronine, about 38% is converted to reverse T3, and about 21% is metabolized via other pathways, such as conjugation in the liver and excretion in the bile. Normal circulating concentrations of thyroxine in plasma range from 4.5 to 11.0 µg/dl, while those of triiodothyronine are about 100-fold less (60 to 180 ng/dl).

The enzyme responsible for the conversion of thyroxine to triiodothyronine is iodothyronine 5'-deiodinase, which exists as two distinct isozymes that are differentially expressed and regulated in peripheral tissues (Figure 56-5; Leonard and Visser, 1986). Type I 5'-deiodinase (5'D-I) is found in the liver, kidney, and thyroid and generates circulating triiodothyronine that is utilized by most peripheral target tissues. Although 5'-deiodination is the major function of this isozyme, 5'D-I also catalyzes 5-deiodination. 5'D-I is inabited by a variety of factors (Table 56-1), including the antithyroid drug, propylthiouracil. The decreased plasma triiodothyronine concentrations observed in nonthyroidal illnesses are a result of inhibition of 5'D-1 (Kaptein, 1986) and decreased entrance of thyroxine into cells. 5'D-I is "up-regulated" in hyperthyroidism and "downregulated" in hypothyroidism. The cloning of 5'D-I has identified the enzyme as a selenoprotein and demonstrated the presence of a selenocystine at the active site (Berry et al., 1991; Berry and Larsen, 1992). Type II 5'-deiodinase (5'D-II) is limited in distribution to the brain, pituitary, and, in the rat, brown fat and functions to supply intracellular triiodothyronine to these tissues (Visser et al., 1982). 5'D-II has a much lower K_m for thyroxine than does 5'D-I (nM vs. μ M

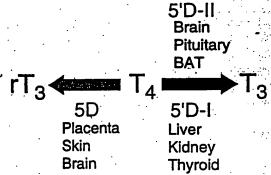


Figure 56-5. Deiodinase isozymes.

Abbreviations are as follows: 5'D-I, type I iodothyronine 5'-deiodinase; 5'D-II, type II iodothyronine 5'-deiodinase; 5D, type III iodothyronine 5-deiodinase; BAT, brown adipose tissue.

Table 56-1
Conditions and Factors That Inhibit Type I
5'-Deiodinase Activity

Acute and chronic illness
Caloric deprivation (especially carbohydrate)
Malnutrition
Glucocorticoids
β-Adrenergic blocking drugs (e.g., propranolol in high doses)
Oral cholecystographic agents (e.g., iopanoic acid, sodium ipodate)
Amiodarone
Propylthiouracil
Fatty acids
Fetal/neonatal period

Selenium deficiency

 K_m values), and its activity is unaffected by propylthiouracil. 5'D-II is dynamically regulated by its substrate, thyroxine, such that elevated levels of the enzyme are found in hypothyroidism and suppressed levels are found in hyperthyroidism (Leonard et al., 1981). Thus, 5'D-II appears to autoregulate the intracellular supply of triodothyronine in the brain and pituitary. 5'D-II is a multimeric protein and is not a selenoenzyme (Safran et al., 1991). Inner ring deiodination, or 5-deiodination, is primarily catalyzed by type III iodothyronine deiodinase (5D), which is found in the placenta, skin, and brain. Whether or not 5D is a selenoprotein is controversial.

Transport of Thyroid Hormones in the Blood. Iodine in the circulation is normally present in several forms, with 95% as organic iodine and approximately 5% as iodide. Most of the organic iodine is thyroxine (90% to 95%), while triiodothyronine represents a relatively minor fraction (about 5%). The thyroid hormones are transported in the blood in strong but noncovalent association with certain plasma proteins.

Thyroxine-binding globulin is the major carrier of thyroid hormones. It is an acidic glycoprotein with a molecular mass of approximately 63 kDa, and it binds one molecule of thyroxine per molecule of protein with a very high affinity (the equilibration association constant, K_o , is about 10^{10} M⁻¹). Triiodothyronine is bound less avidly. Thyroxine, but not triiodothyronine, also is bound by transthyretin (also called thyroxine-binding prealbumin). This protein is present in higher concentration than is the thyroxine-binding globulin, but it binds thyroxine and triiodothyronine with equilibrium association constants near 10^7 M⁻¹ and 10^6 M⁻¹, respectively. Transthyretin has four apparently identical subunits, but has only a single high-affinity binding site. Albumin also can serve as a carrier for thyroxine when the more avid carriers are saturated. It

is difficult, however, to estimate its quantitative or physiological importance, with the exception of the syndrome known as familial dysalbuminemic hyperthyroxinemia. This is an autosomal dominant hereditary disorder characterized by the increased affinity of albumin for thyroxine (Ruiz et al., 1982). Thyroxine binds also to the apolipoproteins of the high density lipoproteins, HDL₂ and HDL₃, the significance of which is unclear at present (Benevenga et al., 1992).

Binding of thyroid hormones to plasma proteins protects the hormones from metabolism and excretion, resulting in their long half-lives in the circulation. The free (unbound) hormone is a small percentage (about 0.03% of thyroxine and about 0.3% of triiodothyronine) of the total hormone in plasma (Larsen et al., 1981). The differential binding affinity for serum proteins also is reflected in the 10- to 100-fold difference in circulating hormone concentrations and half-lives of thyroxine and triiodothyronine.

Essential to understanding the regulation of thyroid function is the "free hormone" concept: only the unbound hormone has metabolic activity (Mendel, 1989). Thus, because of the high degree of binding of thyroid hormones to plasma proteins, changes in either the concentrations of these proteins or the binding affinity of the hormones for the proteins would have major effects on the total serum hormone levels. Certain drugs and a variety of pathological and physiological conditions, such as the changes in circulating concentrations of estrogens during the menstrual cycle, can alter both the binding of thyroid hormones to plasma proteins and the amounts of these proteins (Table 56–2).

Table 56-2

Factors That Alter Binding of Thyroxine to Thyroxine-Binding Globulin

INCREASE BINDING	G DECREASE BINDING				
Drugs					
Estrogens	Glucocorticoids				
Methadone	Androgens				
Clofibrate	L-Asparaginase				
5-Fluorouracil	Salicylates.				
Heroin	Mefenamic Acid				
Tamoxifen	Antiseizure medications (phenytoin, carbamazepine) Furosemide				
Syster	nic Factors				
Liver disease	Inheritance				
Porphyria HIV infection Inheritance	Acute and chronic illness				

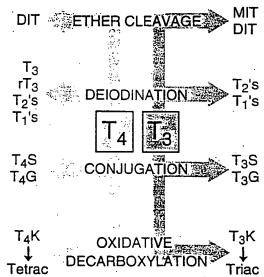


Figure 56-6. Pathways of metabolism of thyroxine (T_4 and triiodothyronine (T_3).

Abbreviations are as follows: DIT, diiodotyrosine; MIT, monoiodotyrosine; T₄S, T₄ sulfate; T₄G, T₄ glucuronide; T₃S, T₃ sulfate; T₃G, T₃ glucuronide; T₄K, T₄ pyruvic acid; T₃K, T₃ pyruvic acid; Tetrac, tetraiodothyroacetic acid. Triac, triiodothyroacetic.

However, since the pituitary responds to and regulates circulating free hormone levels, minimal changes in free hormone concentrations are seen. Laboratory tests that measure only total hormone levels, therefore, can be subject to misinterpretation. Appropriate tests of thyroid function are discussed later in this chapter.

Degradation and Excretion (Figure 56-6). Thyroxine is eliminated slowly from the body, with a half-life of 6 to 7 days. In hyperthyroidism, the half-life is shortened to 3 or 4 days, whereas in hypothyroidism it may be 9 to 10 days. These changes presumably are due to altered rates of metabolism of the hormone. In conditions associated with increased binding to plasma proteins, such as pregnancy, clearance is retarded; the reverse is observed when there is reduced protein binding of thyroid hormones or when binding to protein is inhibited by certain drugs (Table 56-2). Triiodothyronine, which is less avidly bound to protein, has a half-life of approximately 1 day.

The liver is the major site of nondeiodinative degradation of thyroid hormones; thyroxine and triiodothyronine are conjugated with glucuronic and sulfuric acids through the phenolic hydroxyl group and excreted in the bile. There is an enterohepatic circulation of the thyroid hormones; they are liberated by hydrolysis of the conjugates in the intestine and reabsorbed. A portion of the conjugated material reaches the colon unchanged, is hydrolyzed there, and is eliminated

in feces as the free compounds. In human beings, approximately 20% of thyroxine is eliminated in the stool.

As discussed above, the major route of metabolism of thyroxine is deiodination to either triiodothyronine or reverse T₃. Triiodothyronine and reverse T₃ are deiodinated to three different diiodothyronines (see Figure 56-4), inactive metabolites that are normal constituents of human plasma. Additional metabolites (monoiodotyrosine and diiodotyrosine) in which the diphenyl ether linkage is cleaved have been detected both in vitro and in vivo.

Regulation of Thyroid Function. During the past century, it was appreciated that cellular changes occur in the anterior pituitary in association with endemic goiter or following thyroidectomy. The classical experimental observations of Cushing (1912) and the clinical observations of Simmonds (1914) established that ablation or disease of the pituitary causes thyroid hypoplasia. It eventually was determined that thyrotropes of the anterior pituitary secrete thyrotropin, or TSH. TSH is a glycoprotein hormone with α and β subunits analogous to those of the gonadotropins. Its structure is discussed with those of other glycoprotein hormones in Chapter 55. Although there was evidence that thyroid hormone or lack of

causes cellular changes in the pituitary, the control of secretion of TSH by the negative-feedback action of thyroid hormone was not appreciated fully until its central role in the pathogenesis of goiter was elucidated in the early 1940s. TSH is secreted in a pulsatile manner and circadian pattern, its levels in the circulation being highest during sleep at night. It is now recognized that the rate of secretion of TSH is delicately controlled by thyrotropin-releasing hormone (TRH) and the quantity of free thyroid hormones in the circulation. If extra thyroid hormone is given, transcription of the thyrotropin gene is decreased (see Samuels et al., 1988), the secretion of TSH is suppressed, and the thyroid becomes inactive and regresses. Any decrease in the normal rate of secretion of the thyroid evokes an enhanced secretion of TSH in an attempt to stimulate the thyroid to secrete more hormone. Additional mechanisms of the effect of thyroid hormone on TSH secretion appear to be a reduction in TRH secretion by the hypothalamus and a reduction in the number of receptors for TRH on pituitary cells.

Thyrotropin-Releasing Hormone (TRH). TRH stimulates the release of preformed TSH from secretory granules and also stimulates the subsequent synthesis of both α and β subunits of TSH. Somatostatin, dopamine, and pharmacological doses of glucocorticoids inhibit TRH-stimulated TSH secretion.

TRH is a tripeptide with both terminal amino and caruoxyl groups blocked (L-pyroglutamyl-L-histidyl-L-proline amide). The mature hormone is derived from a precursor protein that contains six copies of the tripeptide flanked by dibasic residues. TRH is synthesized by the hypothalamus and is released into the hypophysioportal circulation, where it is brought into contact with TRH receptors on thyrotropes. The binding of TRH to its receptor, a G protein-coupled receptor, elicits stimulation of the hydrolysis of polyphosphatidylinositols and activation of protein kinase C (Gershenghorn, 1986). Ultimately, TRH stimulates the synthesis and release of TSH by the thyrotrope.

TRH also has been localized in the CNS in regions of the cerebral cortex, circumventricular structures, neurohypophysis, pineal gland, and spinal cord. These findings, as well as its localization in nerve endings, suggest that TRH may act as a neurotransmitter or neuromodulator outside of the hypothalamus. Administration of TRH to animals produces CNS mediated effects on behavior, thermoregulation, autonomic tone, and cardiovascular function, including increases in blood pressure and heart rate. TRH also has been identified in pancreatic islet cells and in certain parts of the gastrointestinal tract. Its physiological role there is not known.

Actions of TSH on the Thyroid. When TSH is given to experimental animals, the first effect on thyroid hormone metabolism that can be measured is increased secretion, which can be seen within minutes. All phases of hormone synthesis and release are eventually stimulated: iodide uptake and organification, hormone synthesis, endocytosis, and proteolysis of colloid. There is increased vascularity of the gland and hypertrophy and hyperplasia of thyroid cells. These effects follow the binding of TSH to its receptor on the plasma membrane of thyroid cells.

The TSH receptor is a member of the family of G protein-coupled receptors and is structurally similar to the receptors for luteinizing hormone (LH)-and-follicle-stimulating hormone (FSH) (see Chapter 55; Parmentier et al., 1989; Vassart and Dumont, 1992; Nagayama and Rapoport, 1992). These receptors share significant amino acid sequences and have large extracellular domains that are involved in binding of hormone.

When TSH binds to its receptor, adenylyl cyclase is stimulated and cyclic AMP levels in the cells increase. At higher concentrations than are required to stimulate cyclic AMP formation, TSH causes activation of phospholipase C, with a resultant hydrolysis of polyphosphatidylinositols, increased cytoplasmic Ca²⁺, and activation of protein kinase C (Manley et al., 1988; Van Sande et al., 1990). Both the adenylyl cyclase and the phospholipase C signaling pathways appear to mediate effects of TSH on thyroid function in human beings, although the adenylyl cyclase pathway may be the sole mediating pathway in other species (see Vassart and Dumont, 1992).

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Relation of Iodine to Thyroid Function. Normal thyroid function obviously requires an adequate intake of iodine; without it, normal amounts of hormone cannot be made, TSH is secreted in excess, and the thyroid becomes hyperplastic and hypertrophies. The enlarged and stimulated thyroid becomes remarkably efficient at extracting the residual traces of iodide from the blood. The iodide-concentrating mechanism develops a gradient for the ion that may be ten times normal, and in mild to moderate iodine deficiency, the thyroid usually succeeds in producing sufficient hormone. Adult hypothyroidism and cretinism may occur in more severe iodine deficiency.

In some areas of the world, simple or nontoxic goiter is prevalent because dietary iodine is not sufficient (Delange et al., 1993). Significant regions of iodine deficiency are present in Central and South America, Africa, Europe, southeast Asia, and China. The daily requirement for iodine in adults is 1 to 2 μ g/kg body weight. The United States recommended daily allowance for iodine is in the range of 40 to 120 μ g for children and 150 μ g for adults, with the addition of 25 μ g and 50 μ g recommended during pregnancy and lactation, respectively. Vegetables, meat, and poultry contain minimal amounts of iodine, whereas dairy products and fish are relatively high in iodine content (Table 56–3; Braverman, 1994). Potable water usually contains negligible amounts of iodine.

Iodine has been used empirically for the treatment of iodine-deficiency goiter for 150 years. However, its modern use was the outgrowth of the extensive studies of Marine, which culminated in the use of iodine to prevent goiter in school children in Akron, Ohio, a region where endemic iodine deficiency goiter was prevalent (Marine and Kimball, 1917). The success of these experiments led to the adoption

Table 56–3
Iodine Content in Some Foodstuffs in the United States (1982–1989)

FOOD	1	ODINE/SERVING	μg	
Ready-to-eat cereals	•	87		* ;*
Dairy-based desserts	•	70		· ·
Fish		57		
Milk		, 56 .	_	٠, .
Dairy products		49		
Eggs		27		
Bread		27		
Beans, peas, tuber		17		
Meat	•	16		
Poultry		15		

source: Adapted from Braverman, 1994.

of iodine prophylaxis and therapy in many regions throughout the world where iodine-deficiency goiter is endemic.

The most practicable method for providing small supplements of iodine for large segments of the population is the addition of iodide or iodate to table salt; iodate is now preferred. In some countries, the use of iodized salt is required by law; in others, including the United States, the use is optional. In the United States, iodized salt provides $100~\mu g$ of iodine per gram. Other vehicles for supplying iodine to large populations who are iodine-deficient include oral or intramuscular injection of iodized oil (Thilly et al., 1973), iodized drinking water supplies, iodized irrigation systems, and iodized animal feed.

Actions of Thyroid Hormones. Whereas the precise biochemical mechanisms through which thyroid hormones exert their developmental and tissue-specific effects are only beginning to be understood, the concept that most of the actions of thyroid hormones are mediated by nuclear receptors has been well accepted since the mid-1980s (for review, see Oppenheimer et al., 1987; Brent, 1994). In this model, triiodothyronine binds to high-affinity nuclear receptors, which then bind to a specific DNA sequence (thyroid hormone response element) in the promoter/regulatory region of specific genes. In this fashion, triiodothyronine modulates gene transcription and, ultimately, protein synthesis. In general, the receptor without hormone is bound to the thyroid response element in the basal state. Typically, this results in repressed gene transcription, although there are some examples of constitutive gene activation. Binding by triiodothyronine may activate gene transcription by releasing the repression. Hormone-associated receptors also may have direct activation or repressive actions. Thyroxine also binds to these receptors, but it does so with a much lower affinity than triiodothyronine. It is likely that thyroxine serves principally as a "prohormone." with essentially all actions of thyroid hormone at the transcriptional level being caused by triiodothyronine.

Nuclear thyroid hormone receptors were cloned in 1986 by several laboratories (Weinberger et al., 1986; Sap et al., 1986). They were discovered to be the cellular homologs of an avian retroviral oncoprotein, denoted c-erb A. There is considerable homology between the thyroid hormone receptors and the steroid nuclear receptors, and together they make up a gene superfamily that also includes the retinoic acid and vitamin D nuclear receptors (see Chapters 2 and 63; Mangelsdorf et al., 1994). The thyroid hormone receptors are derived from two genes, c-erb A α (TR α) and c-erb A β (TR β), with multiple isoforms identified (Figure 56–7; Lazar, 1993). TR α 1 and TR β 1 are found in virtually all tissues that respond to thyroid hormone, whereas the other isoforms exhibit a more tissue-specific distribution. TR β 2, for example, is expressed solely in the anterior

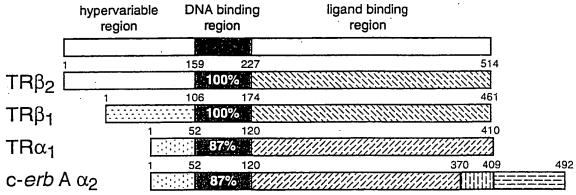


Figure 56-7. Thyroid hormone receptor isoforms.

The percent of amino acid identity in the DNA binding region is indicated. Identical patterns in the hypervariable and ligand binding regions indicate 100% homology. Three thyroid hormone receptor (TR) isoforms bind thyroid hormone (TR β_1 , TR β_2 , and TR α_1); c-erb A α_2 does not.

pituitary. c-erb A α_2 , an isoform that binds to the thyroid response element but does not bind triiodothyronine, is the most abundant isoform in brain (Strait et al., 1990).

In addition to nuclear receptor-mediated actions, there are several well-characterized, nongenomic actions of thyroid hormones, including those occurring at the level of the plasma membrane (Davis et al., 1989) and on the cellular cytoarchitecture (Farwell et al., 1990; Siegrist-Kaiser et al., 1990). In addition, there are well-characterized thyroid hormone binding sites on the mitochondria (Sterling, 1989). In several of these processes, thyroxine is the hormone that produces the response. The overall contribution of the extranuclear sites to cellular regulation by thyroid hormones is likely to be minor.

Growth and Development. As discussed above, it is generally believed that the thyroid hormones exert most if not all of their effects through control of DNA transcription and, ultimately, protein synthesis. This is certainly true for the actions of the hormones on the normal growth and development of the organism. Perhaps the most dramatic example is found in the tadpole, which is almost magically transformed into a frog by thyroid hormone. Not only does the animal grow limbs, lungs, and other terrestrial accoutrements, but the hormone also stimulates the synthesis of a host of enzymes and so influences the tail that it is digested away and used to build new tissue elsewhere.

Thyroid hormone plays a critical role in brain development (Dussault and Ruel, 1987; Porterfield and Hendrich, 1993). The appearance of functional, chromatin-bound receptors for thyroid hormone coincides with neurogenesis in the brain (Strait et al., 1990). The absence of thyroid hormone during the period of active neurogenesis (up to 6 months postpartum) leads to irreversible mental retardation (cretinism) and is accompanied by multiple morphological alterations in the brain (Legrand, 1979). These severe morphological alterations result from disturbed neuronal migration, deranged axonal projections,

and decreased synaptogenesis. Thyroid hormone supplementation during the first 2 weeks of life prevents the development of these disturbed morphological changes.

Myelin basic protein, a major component of myelin, is the product of a specific gene that is regulated by thyroid hormone during development (Farsetti et al., 1991). Decreased expression of myelin basic protein results in defective myelinization in the hypothyroid brain. Several other minor brain-specific genes have been reported to be developmentally regulated by thyroid hormone (Porterfield and Hendrich, 1993). A common characteristic of these proteins is that their expression appears to be merely delayed in the hypothyroid animal; normal levels are eventually achieved in the adult.

The actions of thyroid hormones on protein synthesis and enzymatic activity are certainly not limited to the brain, and a large number of tissues are affected by the administration of thyroid hormone or by its deficiency. The extensive defects in growth and development that are found in cretins provide a vivid reminder of the pervasive effects of thyroid hormones in normal individuals.

Cretinism is usually classified as endemic or sporadic. Endemic cretinism is encountered in regions of endemic goiter and is usually caused by extreme deficiency of iodine. Goiter may or may not be present. Sporadic cretinism is a consequence of failure of the thyroid to develop normally or the result of a defect in the synthesis of thyroid hormone. Goiter is present if a synthetic defect is at fault.

While detectable at birth, cretinism often is not recognized until 3 to 5 months of age. When untreated, the condition eventually leads to such gross changes as to be unmistakable. The child is dwarfed, with short extremities, and is mentally retarded, inactive, uncomplaining, and listless. The face is puffy and expressionless, and the enlarged tongue may prorude through the thickened lips of the half-opened mouth. The skin may have a yellowish hue and feel doughy, and it is dry and cool to the touch. The heart rate is slow, the body temperature may be low, closure of the fontanels is delayed, and the teeth erupt late. Appetite is poor, feeding is slow and interrupted by choking, constipation is frequent, and there may be an umbilical hernia.

For treatment to be fully effective, the diagnosis must be made long before these obvious changes have come about. Screening of newborn infants for deficient function of the thyroid is carried out in the United States and in most industrialized countries. Concentrations of TSH and thyroxine are measured in blood from the umbilical cord or from a heel stick. The incidence of congenital dysfunction of the thyroid is about 1 per 4000 births (Fisher, 1991).

Calorigenic Effect. A characteristic response of homeothermic animals to thyroid hormone is increased oxygen consumption (Oppenheimer, 1991). Most peripheral tissues contribute to this response; heart, skeletal muscle, liver, and kidney are stimulated markedly by thyroid hormone. Indeed, 30% to 40% of the thyroid hormone-dependent increase in oxygen consumption can be attributed to stimulation of cardiac contractility. Several organs, including brain, gonads, and spleen, are unresponsive to the calorigenic effects of thyroid hormone. The mechanism of the calorigenic effect of thyroid hormone has been elusive. At one time, it was erroneously believed that thyroid hormone uncoupled mitochondrial oxidative phosphorylation. Thyroid hormonedependent lipogenesis may constitute a quantitatively important energy sink, and studies in rats have demonstrated that about 4% of the increased caloric expenditure induced by thyroid hormone is accounted for by lipogenesis. A link between lipogenesis and thermogenesis is the stimulation of lipolysis by triiodothyronine. Further, thyroid hormone induces expression of several lipogenic enzymes, including malic enzyme and fatty acid synthetase. Although the entire picture is not clear, there appears to be an integrated thyroid hormone response program for regulating the set-point of energy expenditure and maintaining the metabolic machinery necessary to sustain it.

Cardiovascular Effects. Thyroid hormone influences cardiac function by direct and indirect actions; changes in the cardiovascular system are prominent clinical consequences in thyroid dysfunctional states. In hyperthyroidism, there is tachycardia, increased stroke volume, increased cardiac index, cardiac hypertrophy, decreased peripheral vascular resistance, and increased pulse pressure. In hypothyroidism, there is bradycardia, decreased cardiac index, pericardial effusion, increased peripheral vascular resistance, decreased pulse pressure, and elevations of mean arterial pressure. (For a review of the effects of thyroid hormone on the heart, see Braverman et al., 1994.)

Thyroid hormones play a direct role in regulating myocardial gene expression. Triiodothyronine regulates genes encoding the isoforms of the sarcomeric myosin heavy chains by increasing the expression of the α gene and decreasing the expression of the β gene (Everett et al., 1986). A thyroid hormone response element has been located in the 5' upstream region of the α myosin heavy chain gene. Triiodothyronine also upregulates the gene encoding myosin Ca^{2+} -ATPase, which plays a critical role in myocardial contraction

(Rohrer and Dillman, 1989). Regulation of these two genes results in the changes in contractility observed in hyper- and hypothyroidism.

Thyroid hormones also indirectly influence cardiac function. The sensitivity of the cardiac myocyte to catecholamines is enhanced in hyperthyroidism and depressed in hypothyroidism, possibly due to changes in expression of myocardial β -adrenergic receptors; this is the basis for the use of β -adrenergic receptor antagonists in relieving some of the cardiac manifestations of hyperthyroidism. Electrical impulse generation and conduction are increased in hyperthyroidism and decreased in hypothyroidism. These two actions likely account for the chronotropic effects of triiodothyronine. Finally, triiodothyronine causes hemodynamic alterations in the periphery that result in alterations in the chronotropic and ionotropic state of the myocardium.

Metabolic Effects. Thyroid hormones stimulate metabolism of cholesterol to bile acids, and hypercholesterolemia is a characteristic feature of hypothyroid states. Thyroid hormones have been shown to increase the specific binding of low density lipoprotein (LDL) by liver cells (Salter et al., 1988), and the concentration of hepatic receptors for LDL is decreased in hypothyroidism (Scarabottolo et al., 1986; Gross et al., 1987). The number of LDL receptors available on the surface of hepatocytes is a strong determinant of the plasma cholesterol concentration (see Chapter 36).

Thyroid hormones enhance the lipolytic responses of fat cells to other hormones, for example, catecholamines, and elevated plasma free fatty acid concentrations are seen in hyperthyroidism. In contrast to other lipolytic hormones, thyroid hormones do not directly increase the accumulation of cyclic AMP. They may, however, regulate the capacity of other hormones to enhance the accumulation of the cyclic nucleotide by decreasing the activity of a microsomal phosphodiesterase that hydrolyzes cyclic AMP (Nunez and Correze, 1981). There also is evidence that thyroid hormones act to maintain normal coupling of the β -adrenergic receptor to the catalytic subunit of adenylyl cyclase in fat cells. Fat cells from hypothyroid rats have increased concentrations of guanine nucleotide-binding regulatory proteins (G proteins) that mediate the inhibitory control of adenylyl cyclase (see Chapter 2). This can account for both the decreased response to lipolytic hormones and the increased sensitivity to inhibitory regulators, such as adenosine, that are found in hypothyroidism (Ros et al., 1988).

Thyrotoxicosis is an insulin-resistant state (Gottlieb and Braverman, 1994). Postreceptor defects in the liver and peripheral tissues, manifested by depleted glycogen stores and enhanced glucogenesis, lead to insulin insensitivity. In addition, there is increased absorption of glucose from the gut. Compensatory increases in insulin secretion result in order to maintain euglycemia. This may result in the "unmasking" of clinical diabetes in previously undiagnosed patients and an increase in the insulin requirements of diabetic patients already on insulin. Hypothyroidism results in decreased absorption of glucose from the gut and decreased insulin secretion. Peripheral glucose uptake also is slowed in hypothyroidism, although glucose utilization by the brain is unaffected. Insulin requirements are decreased in the hypothyroid patient with diabetes.

Thyroid Hyperfunction. Thyrotoxicosis is a condition caused by elevated concentrations of circulating free thyroid hormones. Various disorders of different etiologies can result in this syndrome. The term hyperthyroidism is restricted to those conditions in which thyroid hormones are excessively released due to gland hyperfunction. Iodine up-

take by the thyroid gland is increased, as determined by the measurement of the percent uptake of ¹²³I or ¹³¹I in a 24-hour radioactive iodine uptake (RAIU) test. In contrast, thyroid inflammation or destruction resulting in excess "leak" of thyroid hormones or exogeneous thyroid hormone intake results in a low 24-hour RAIU.

Graves' disease, or toxic diffuse goiter, is the most common cause of high RAIU thyrotoxicosis. It accounts for 60% to 90% of the cases, depending upon age and geographic region. Graves' disease is an autoimmune disorder characterized by hyperthyroidism, diffuse goiter, and IgG antibodies that bind to and activate the TSH receptor (Burman and Baker, 1985; Bottazzo and Doniach, 1986). This is a relatively common disorder, with an incidence of 0.02% to 0.4% in the United States. Endemic areas of iodine deficiency have a lower incidence of autoimmune thyroid disease. As with most types of thyroid dysfunction, women are affected more than men, with a ratio ranging from 5:1 to 7:1. Graves' disease is more common between the ages of 20 and 50, but may occur at any age. HLA Bg and DR₃ haplotypes are associated with Graves' disease in Caucasians. Graves' disease is commonly associated with other autoimmune diseases. The characteristic exophthalmos associated with Graves' disease is an infiltrative ophthalmopathy and is considered an autoimmune-mediated inflammation of the periorbital connective tissue and extraocular muscle. This disorder is clinically evident with various degrees of severity in about 50% of patients with Graves' isease, but is present on radiological studies, such as ultrasound or T scan, in almost all patients. Two reviews on the pathogenesis and management of Graves' ophthalmopathy recently have been published (Burch and Wartofsky, 1993; Bahn and Heufelder, 1993).

Toxic uninodular and multinodular goiter accounts for 10% to 40% of cases of hyperthyroidism and is more common in older patients. Infiltrative ophthalmopathy is absent.

A low RAIU is seen in the destructive thyroiditides and in thyrotoxicosis resulting from exogenous thyroid hormone ingestion. Low RAIU thyrotoxicosis caused by subacute (painful) and silent (painless or lymphocytic) thyroiditis represents about 5% to 20% of all cases. Silent thyroiditis occurs in 7% to 10% of postpartum women in the United States (Roti and Emerson, 1992). Other causes of thyrotoxicosis are much less common.

Most of the signs and symptoms of thyrotoxicosis stem from the excessive production of heat and from increased motor activity and increased activity of the sympathetic nervous system. The skin is flushed, warm, and moist; the muscles are weak and tremulous; the heart rate is rapid, and the heart beat is forceful; and the arterial pulses are prominent and bounding. The increased expenditure of energy gives rise to increased appetite and, if intake is insufficient, to loss of weight. There also may be insomnia, difficulty in remaining still, anxiety and apprehension, intolerance to heat, and increased frequency of bowel movements. Angina, arrhythmias, and heart failure may be present in older patients. Some individuals may show extensive muscular wasting as a result of thyroid myopathy. Patients with long-standing undiagnosed or undertreated thyrotoxicosis may develop osteoporosis due to increased bone turnover (Baran, 1994).

Thyroid Hypofunction. Hypothyroidism, known as yxedema when severe, is the most common disorder of thyroid function. Worldwide, hypothyroidism is most often the result of iodine deficiency. In nonendemic areas,

where iodine is sufficient, chronic autoimmune thyroiditis (Hashimoto's thyroiditis) accounts for the majority of cases. Failure of the thyroid to produce sufficient thyroid hormone is the most common cause of hypothyroidism and is referred to as primary hypothyroidism. Central hypothyroidism occurs much less often and results from diminished stimulation of the thyroid by TSH because of pituitary failure (secondary hypothyroidism) or hypothalamic failure (secondary hypothyroidism). Hypothyroidism present at birth is known as congenital hypothyroidism and is the most common preventable cause of mental retardation in the world. Diagnosis and early intervention with thyroid hormone replacement prevent the development of cretinism, as discussed above.

Nongoitrous hypothyroidism is associated with degeneration and atrophy of the thyroid gland. The same condition follows surgical removal of the thyroid or its destruction by radioactive iodine. Since it also may occur years after antithyroid drug therapy for Graves' disease, some have speculated that hypothyroidism can be the end stage of this disorder ("burnt-out" Graves' disease). Goitrous hypothyroidism occurs in Hashimoto's thyroiditis and when there is a severe defect in synthesis of thyroid hormone. When the disease is mild, it may be subtle in its presentation. By the time it has become severe, however, all of the signs are overt. The appearance of the patient is pathognomonic. The face is quite expressionless, puffy, and pallid. The skin is cold and dry, the scalp is scaly, and the hair is coarse, brittle, and sparse. The fingernails are thickened and brittle, the subcutaneous tissue appears to be thickened, and there may be true edema. The voice is husky and low-pitched, speech is slow, hearing is often faulty, and mentation is impaired and depression may be present. The appetite is poor, gastrointestinal activity is diminished, and constipation is common. Atony of the bladder is rare and suggests that the function of other smooth muscles may be impaired. The voluntary muscles are weak and the relaxation phase of the deep-tendon reflexes is delayed. The heart can be dilated, and there is frequently a pericardial effusion, although this is rarely clinically significant. There also may be pleural effusions and ascites. Anemia most commonly normochromic, normocytic, is often present, although menstrual irregularity with menorrhagia may result in iron deficiency anemia. Patients are lethargic and tend to sleep a lot and often complain of cold intolerance.

Thyroid Function Tests. The development of radioimmunoassays and, more recently, chemiluminescent and enzyme-linked immunoassays for thyroid hormones have greatly improved the laboratory diagnosis of thyroid disorders (Surks et al., 1990). However, measurement of the total hormone concentration in plasma may not give an accurate picture of the activity of the thyroid gland. The total hormone concentration changes with alterations in either the amount of thyroxine-binding globulin (TBG) or the binding affinity for hormones to TBG in plasma. Although equilibrium dialysis of undiluted serum and radioimmunoassay for free thyroxine in the dialysate represent the gold standard for determining free thyroxine concentrations, this assay is labor-intensive and typically not available in routine clinical laboratories (Nelson and Tomei, 1988). The free thyroxine index is an estimation of the free thyroxine concentration and is calculated by multiplying the total thyroxine concentra-

tion by the thyroid hormone binding ratio, which estimates the degree of saturation of TBG (Nelson and Tomei, 1989). Additional procedures for estimating free thyroxine levels include radioimmunoassay using radiolabeled analogs of thyroxine that do not perturb the thyroxine—thyroid-binding globulin as the tracer (Nelson and Weiss, 1985), and a sequential thyroxine-binding/radiolabeled thyroxine competition assay, dubbed the two-step T₄ assay. This assay correlates well with free thyroxine levels measured by the more cumbersome equilibrium dialysis determination of thyroxine concentration, yet is easily adaptable to routine clinical laboratory use (Wilke, 1986).

Estimates of free thyroxine levels should be complemented with serum measurements of TSH. In fact, in individuals whose pituitary function and TSH secretion are normal, serum measurements of TSH may be the thyroid function test of choice, because pituitary secretion of TSH is sensitively regulated in response to circulating concentrations of thyroid hormones. The American Thyroid Association has published a report outlining for clinicians a suggested approach using a limited number of tests in the laboratory diagnosis of thyroid disorders, suggesting the estimates of free thyroxine and a sensitive TSH assay (Surks et al., 1990).

Serum measurements of TSH have been available since 1965 and have become the thyroid function test of choice. The first assays were single antibody radioimmunoassays and remained the standard for 20 years. These assays were useful only for diagnosing primary hypothyroidism, as a lower limit of the normal range could not be reliably measured. The first "sensitive" TSH assay was developed in 1985, utilizing a dual antibody approach. Application of this method resulted in the expansion of the assay detection limit below the normal range. Thus, any assay of this type is referred to as a sensitive TSH assay (Nicoloff and Spencer, 1990). A major use of the sensitive TSH assay is to differentiate between normal and thyrotoxic patients, who should exhibit suppressed TSH values. The response of TSH to an injection of synthetic TRH (TRH stimulation test) may be useful in determining pituitary or hypothalamic failure as a cause of hypothyroidism. Synthetic preparations of thyrotropin-releasing hormone (protirelin, RELEFACT TRH, THYPINONE) are available for injection for the TRH stimulation test.

Thyrotropin (THYTROPAR) was available as an injectable preparation made from bovine pituitaries. It is no longer available because of the high incidence of anaphylaxis. This preparation was used to test the ability of thyroid tissue to take up radioactive iodine. Recombinant human TSH (THYROGEN) soon will be available for this use (Meier et al., 1994).

Therapeutic Uses of Thyroid Hormone. The major indications for the therapeutic use of thyroid hormone are for hormone replacement therapy in patients with hypothyroidism or cretinism and for TSH suppression therapy in patients with nontoxic goiter or after treatment for thyroid cancer (Roti et al., 1993; Toft, 1994). Thyroid hormone therapy is not indicated for treatment of the "low T₄ syndrome" ("sick euthryoid syndrome") that is a result of nonthyroidal illness (Brent and Hershman, 1986).

The synthetic preparations of the sodium salts of the natural isomers of the thyroid hormones are available and widely used for thyroid hormone therapy. Levothyroxine sodium (L-T₄, SYNTHROID, LEVOTHROID, others) is available in tablets and as a lyophilized powder for injection. Liothyronine sodium (L-T₃) is the salt of tri-iodothyronine and is available in tablets (CYTOMEL) and in an in-

jectable form (TRIOSTAT). A mixture of thyroxine and triiodothyronine is marketed as *liotrix* (THYROLAR). Desiccated thyroid preparations, derived from whole animal thyroids, contain both thyroxine and triiodothyronine and have highly variable biologic activity, making these preparations much less desirable.

Thyroid Hormone Replacement Therapy. Thyroxine (levothyroxine sodium) is the hormone of choice for thyroid hormone replacement therapy because of its consistent potency and prolonged duration of action. The absorption of thyroxine occurs in the small intestine and is variable and incomplete, with 50% to 80% of the dose absorbed (Hays, 1991; Hays and Nielson, 1994). Absorption is increased when the hormone is taken on an empty stomach. In addition, certain drugs may interfere with absorption of levothyroxine in the gut, including sucralfate, cholestyramine resin, iron supplements, and aluminum hydroxide. Triiodothyronine (liothyronine sodium) may be used occasionally when a quicker onset of action is desired, as, for example, in the rare presentation of myxedema coma or for preparing a patient for ¹³¹I therapy for treatment of thyroid cancer. It is less desirable for chronic replacement therapy because of the requirement for more frequent dosing, higher cost, and transient elevations of serum triiodothyronine concentrations above the normal range.

The average daily adult replacement dose of levothyroxine sodium in a 68-kg person is 112 μ g. That of liothyronine sodium is 25 to 50 μ g. Institution of therapy in healthy younger individuals can begin at full replacement doses. Because of the prolonged half-life of thyroxine (7 days), new steady-state concentrations of the hormone will not be achieved until 4 to 6 weeks after a change in dose. Thus, reevaluation with determination of serum TSH concentration need not be performed at intervals less than 4 to 6 weeks. The goal of thyroxine replacement therapy is to achieve a TSH value in the normal range, as overreplacement of thyroxine suppressing TSH values to the subnormal range may induce osteoporosis and cause cardiac dysfunction (Ross, 1991). In individuals over the age of 60, institution of therapy at a lower daily dose of levothyroxine sodium (25 µg per day) is indicated to avoid exacerbation of underlying and undiagnosed cardiac disease. Death due to arrhythmias has been reported during the initiation of thyroid hormone replacement therapy in hypothyroid patients. The dose can be increased at a rate of 25 μ g per day every few months until the TSH is normalized. For individuals with preexisting cardiac disease, an initial dose of 12.5 µg per day, with increases of 12.5 to 25 μ g per day every 6 to 8 weeks, is indicated. Daily doses of thyroxine may be interrupted periodically because of intercurrent medical or surgical illnesses that prohibit taking medications by mouth. A lapse of several days of hormone replacement is unlikely to have any significant metabolic consequences. However, if more prolonged interruption in oral therapy is necessary, levothyroxine may be given parenterally at a dose 25% to 50% less than the patient's daily oral requirements.

The decision to use levothyroxine therapy in patients with elevated serum TSH values but serum thyroxine and triiodothyronine concentrations in the normal range, a syndrome known as subclinical hypothyroidism, must be made on an individual basis (Cooper, 1991a). Patients with subclinical hypothyroidism and goiter, autioimmune thyroid disease, hypercholesterolemia, or symptoms of hypothyroidism may benefit from levothyroxine therapy.

The dose of levothyroxine in the hypothyroid patient who becomes pregnant often needs to be increased, perhaps due to the increased serum concentrations of thyroid-binding globulin induced by estrogen (Kaplan, 1992; Glinoer, 1993). In addition, pregnancy may "unmask" hypothyroidism in patients with preexisting autoimmune

thyroid disease or in those who reside in a region of iodine deficiency (Glinoer et al., 1994). Thus, serum TSH values should be determined in the first trimester in these patients and followed each trimester in patients with documented hypothyroidism, and the levothyroxine dose adjusted to keep the serum TSH in the normal range.

Comparative Responses to Thyroid Preparations. There is no significant difference in the qualitative response of the patient with myxedema to triiodothyronine, thyroxine, or desiccated thyroid. However, there are obvious quantitative differences. Following the subcutaneous administration of a large experimental dose of triiodothyronine, a metabolic response can be detected within 4 to 6 hours, at which time the skin becomes detectably warmer and the pulse rate and temperature increase. With this dose, a 40% decrease in metabolic rate can be restored to normal in 24 hours. The maximal response occurs in 2 days or less, and the effects subside with a halftime of about 8 days. The same single dose of thyroxine exerts much less effect. However, if thyroxine is given in approximately four times the dose of triiodothyronine, a comparable elevation in metabolic rate can be achieved. The peak effect of a single dose is evident in about 9 days, and this declines to half the maximum in 11 to 15 days. In both cases the effects outlast the presence of detectable amounts of hormone; these disappear from the blood with mean half-lives of approximately 1 day for triiodothyronine and 7 days for thyroxine.

Myxedema Coma. Myxedema coma is a rare syndrome that represents the extreme expression of severe, long-standing hypothyroidism (Gavin, 1991; Smallridge, 1992). It is a medical emergency, and even with early diagnosis and treatment, the mortality rate can be as high 60%. Myxedema coma occurs most often in elderly patients dur-

g the winter months. Common precipitating factors include pulmonary infections, cerebrovascular accidents, and congestive heart failure. The clinical course of lethargy proceeding to stupor and then coma is often hastened by drugs, especially sedatives, narcotics, antidepressants, and tranquilizers. Indeed, many cases of myxedema coma have occurred in hypothyroid patients who have been hospitalized for other medical problems.

Cardinal features of myxedema coma are: (1) hypothermia, which may be profound, (2) respiratory depression, and (3) unconsciousness. Other clinical features include bradycardia, macroglossia, delayed reflexes, and dry, rough skin. Dilutional hyponatremia is common and may be severe. Elevated plasma creatine phosphokinase (CPK) and lactate dehydrogenase (LDH) concentrations, acidosis and anemia are common findings. Lumbar puncture reveals increased opening pressure and high protein content. Hypothyroidism is confirmed by measuring serum free thyroxine index and TSH values. Ultimately, myxedema coma is a clinical diagnosis.

The mainstay of therapy is supportive care, with ventilatory support, rewarming, correction of hyponatremia, and treatment of the precipitating incident. Because of a 5% to 10% incidence of coexisting decreased adrenal reserve in patients with myxedema coma, intravenous steroids are indicated before initiating thyroxine therapy. Parenteral administration of thyroid hormone is necessary due to uncertain absorption through the gut. With intravenous preparations of both levothyroxine and liothyronine now available, a reasonable approach is an initial intravenous loading dose of 200 to 300 μ g of levothyroxine with a second dose of 100 μ g given 24 hours later. Simultaneously with the initial dose of levothyroxine, some clinicians recommend adding liothyronine at a dose of 10 μ g intravenously every 8 hours until the patient is stable and conscious. The dose of

roid hormone should be adjusted on the basis of hemodynamic ability, the presence of coexisting cardiac disease, and the degree of electrolyte imbalance.

Treatment of Cretinism. Success in the treatment of cretinism depends upon the age at which therapy is started. Because of this, newborn screening for congenital hypothyroidism is routine in the United States, Canada, and many other countries around the world. In cases that do not come to the attention of physicians until retardation of development is clinically obvious, the detrimental effects of thyroid hormone deficiency on mental development will not be overcome. If, on the other hand, therapy is instituted within the first few weeks of life, normal physical and mental development is almost always achieved. Prognosis also depends on the severity of the hypothyroidism at birth and may be worse for babies with thyroid agenesis. The most critical need for thyroid hormone is during the period of myelinization of the central nervous system that occurs about the time of birth. To rapidly normalize the serum thyroxine concentration in the congenitally hypothyroid infant, an initial daily dose of levothyroxine of 10 to 15 μ g/kg is recommended (Fisher, 1991). This dose will increase the total serum thyroxine concentration to the upper half of the normal range in most infants within 1 to 2 weeks. Individual levothyroxine doses are adjusted at 4- to 6-week intervals during the first 6 months, at 2month intervals during the 6- to 18-month period, and at 3- to 6-month intervals thereafter to maintain serum thyroxine concentrations in the 10 to 16 μ g/dl range and serum TSH values below 20 mU/l. The free thyroxine levels should be kept in the upper normal or elevated range. Assessments that are important guides for appropriate hormone replacement include physical growth, motor development, bone maturation, and developmental progress.

Nodular Thyroid Disease. Nodular thyroid disease is the most common endocrinopathy. The prevalence of clinically apparent nodules is 4% to 7% in the United States, with the frequency increasing throughout adult life. When ultrasound and autopsy data are included, the prevalence of thyroid nodules approaches 50% by age 60. As with other forms of thyroid disease, nodules are more frequent in women. Nodules have been estimated to develop at a rate of 0.1% per year. In individuals exposed to ionizing radiation, the rate of nodule development is 20-fold higher. While the presence of a nodule raises the question of a malignancy, only 8% to 10% of patients with thyroid nodules have thyroid cancer. About 12,000 new cases of thyroid cancer are diagnosed annually, with about 1000 deaths from the disease per year. However, many more people have clinically silent thyroid cancer, as up to 35% of thyroids removed at autopsy or at surgery harbor a small (<1 cm) occult papillary cancer.

The evaluation of the patient with nodular thyroid disease includes a careful physical examination, biochemical analysis of thyroid function, and assessment of the malignant potential of the nodule (Mazzaferri, 1993; Gharib and Goellner, 1993). The latter often includes examination of a fine-needle aspiration biopsy of the nodule and radioisotope scanning with 123I or 131I to determine if a particular nodule is functioning. TSH suppressive therapy with levothyroxine is an option for the patient diagnosed with a benign solitary nodule. The rationale behind levothyroxine therapy is that the benign nodule will either stop growing or decrease in size after TSH stimulation of the thyroid gland has been suppressed. The success rate of such therapy ranges from 0 to 68% in different studies. Identification of those patients who are most likely to benefit from thyroid hormone therapy can be achieved through measurement of the serum TSH concentration and radioisotope scanning. Suppression therapy will be of no value if thyroid nodule autonomy exists, as evidenced by a subnormal TSH value and all isotope uptake in the nodule. Functioning nodules are the most likely to respond to suppression therapy. However, once TSH concentrations are suppressed, a repeat radioisotope scan (suppression scan) should be obtained. If significant

uptake persists on a suppression scan, the nodule is nonsuppressible and levothyroxine therapy should be discontinued. Suppression therapy needs to be considered carefully in older patients or in those with coronary artery disease, and, in general, such therapy should be avoided in these patients. Hypofunctioning nodules are much less likely to respond to suppression therapy. However, a 6- to 12-month trial of levothyroxine suppression is reasonable. If levothyroxine is administered, therapy should be continued for as long as the nodule is decreasing in size. Once the size of a nodule remains stable for a 6- to 12-month period, therapy may be discontinued and the nodule observed for recurrent growth. Any nodule that grows while on suppression therapy should be rebiopsied and/or surgically excised.

ANTITHYROID DRUGS AND OTHER THYROID INHIBITORS

A large number of compounds are capable of interfering, directly or indirectly, with the synthesis, release, or action of thyroid hormones (Table 56-4). Several are of great clinical value for the temporary or extended control of hyperthyroid states. These will be discussed in detail. Others are primarily of research or toxicological interest and are only mentioned briefly. The major inhibitors may be classified

into four categories: (1) antithyroid drugs, which interfere directly with the synthesis of thyroid hormones; (2) ionic inhibitors, which block the iodide transport mechanism; (3) high concentrations of iodine itself, which decrease release of thyroid hormones from the gland and also may decrease hormone synthesis; and (4) radioactive iodine, which damages the gland with ionizing radiation. Adjuvant therapy with drugs that have no specific effects on thyroid gland hormonogenesis is useful in controlling the peripheral manifestations of thyrotoxicosis. These drugs include: inhibitors of the peripheral deiodination of thyroxine to the active hormone, triiodothyronine; β -adrenergic receptor antagonists; and Ca2+ channel blockers. The antithyroid drugs have been reviewed by Green (1991) and Cooper (1984). Adrenergic agents are discussed more fully in Chapter 10 and Ca²⁺ channel blockers in Chapter 35.

Antithyroid Drugs

The antithyroid drugs that have clinical utility are the thioureylenes, which belong to the family of thionamides. Propylthiouracil may be considered as the prototype.

Table 56-4
Antithyroid Compounds

PROCESS AFFECTED	EXAMPLES OF INHIBITORS
Active transport of iodide	Complex anions: perchlorate, fluoborate, pertechnetate, thiocyanate
Iodination of thyroglobulin	Thionamides: propylthiouracil, methimazole, carbimazole
•	Thiocyanate
	Aniline derivatives; sulfonamides
	Iodide
Coupling reaction	Thionamides
	Sulfonamides
	?All other inhibitors of iodination
Hormone release	Lithium salts
en e	Iodide
Iodotyrosine deiodination	Nitrotyrosines
Peripheral iodothyronine	Thiouracil derivatives
deiodination	Oral cholecystographic agents
•	Amiodarone
Hormone excretion/ inactivation	Inducers of hepatic drug-metabolizing enzymes: phenobarbital, rifampin, carbamazepine, phenytoin
Hormone action	Thyroxine analogs Amiodarone ?Phenytoin

SOURCE: Adapted from Green, 1991.

History. Studies on the mechanism of the development of goiter began with the observation that rabbits fed a diet composed largely of cabbage often developed goiters. This result was probably due to the presence of precursors of the thiocyanate ion in cabbage leaves (see below). Later, two pure compounds were shown to produce goiter, sulfaguanidine and phenylthiourea.

Investigation of the effects of thiourea derivatives revealed that rats became hypothyroid despite hyperplastic changes in their thyroid glands that were characteristic of intense thyrotropic stimulation. After treatment was begun, no new hormone was made, and the goitrogen had no visible effect upon the thyroid gland following hypophysectomy or the administration of thyroid hormone. This suggested that the goiter was a compensatory change resulting from the induced state of hypothyroidism and that the primary action of the compounds was to inhibit the formation of thyroid hormone (Astwood, 1945). The therapeutic possibilities of such agents in hyperthyroidism were evident, and the substances so used became known as antithyroid drugs.

Structure-Activity Relationship. The two goitrogens found in the early 1940s proved to be prototypes of two different classes of antithyroid drugs. These two, with one later addition, made up three

neral categories into which the majority of the agents can be assigned: (1) thioureylenes include all the compounds currently used clinically (Figure 56-8); (2) aniline derivatives, of which the sulfonamides make up the largest number, embrace a few substances that have been found to inhibit thyroid hormone synthesis; and (3) polyhydric phenols, such as resorcinol, which have caused goiter in human beings when applied to the abraded skin. A few other compounds, mentioned briefly below, do not fit into any of these categories.

Thiourea and its simpler aliphatic derivatives and heterocyclic compounds containing a thioureylene group make up the majority of the known antithyroid agents that are effective in human beings. Although most of them incorporate the entire thioureylene group, in some a nitrogen atom is replaced by oxygen or sulfur so that only the thioamide group is common to all. Among the heterocyclic compounds, active representatives are the sulfur derivatives of imidazole, oxazole, hydantoin, thiazole, thiadiazole, uracil, and barbituric acid.

L-5-Vinyl-2-thiooxazolidone (goitrin) is responsible for the goiter that results from consuming turnips or the seeds or green parts of cruciferous plants. These plants are eaten by cows, and the compound is found in cow's milk in areas of endemic goiter in Finland; it is about as active as propylthiouracil in human beings. VanEtten (1969) has reviewed the chemistry of naturally occurring goitrogens.

Figure 56-8. Antithyroid drugs of the thiamide type.

As the result of industrial exposure, toxicological studies, or clinical trials for various purposes, several other compounds have been noted to possess antithyroid activity (Gaitan, 1989; McKinney and Waller, 1994). Thiopental and oral hypoglycemic drugs of the sulfonylurea class have weak antithyroid action in experimental animals. This is not significant at usual doses in human beings. However, antithyroid effects in human beings have been observed from dimercaprol, aminoglutethimide, and lithium salts. Amiodarone, the iodine-rich drug used in the management of cardiac arrhythmias, has complex effects on thyroid function (Gammage and Franklyn, 1987). In areas of iodine sufficiency, amiodarone-induced hypothyroidism is not uncommon, whereas in iodine-deficient regions, amiodaroneinduced thyrotoxicosis predominates, whether because of the excess iodine or the thyroiditis induced by the drug. Amiodarone is a potent inhibitor of iodothyronine deiodination, resulting in decreased conversion of thyroxine to triiodothyronine. In addition, its major metabolite, desmethylamiodarone, decreases binding of triiodothyronine to its nuclear receptors.

Mechanism of Action. The mechanism of action of the thiourylene drugs has been thoroughly discussed by Taurog (1991). Antithyroid drugs inhibit the formation of thyroid hormones by interfering with the incorporation of iodine into tyrosyl residues of thyroglobulin; they also inhibit the coupling of these iodotyrosyl residues to form iodothyronines. This implies that they interfere with the oxidation of iodide ion and iodotyrosyl groups. Taurog (1976) proposed that the drugs inhibit the peroxidase enzyme, thereby preventing oxidation of iodide or iodotyrosyl groups to the required active state. Subsequent studies have confirmed that this is, indeed, the mechanism of action and that the antithyroid drugs bind to and inactivate the peroxidase only when the heme of the enzyme is in the oxidized state (Davidson et al., 1978; Engler et al., 1982). Over a period of time, the inhibition of hormone synthesis results in the depletion of stores of iodinated thyroglobulin as the protein is hydrolyzed and the hormones are released into the circulation. Only when the preformed hormone is depleted and the concentrations of circulating thyroid hormones begin to decline do clinical effects become noticeable.

There is some evidence that the coupling reaction may be more sensitive to an antithyroid drug, such as propylthiouracil, than is the iodination reaction (Taurog, 1991). This may explain why patients with hyperthyroidism respond well to doses of the drug that only partially suppress organification.

When Graves' disease is treated with antithyroid drugs, the concentration of thyroid-stimulating immunoglobulins in the circulation often decreases. This has prompted some to propose that these agents act as immunosuppressants. Burman and Baker (1985) point out that perchlorate, which acts by an entirely different mechanism, also decreases thyroid-stimulating immunoglobu-

lins, suggesting that improvement in hyperthyroidism may, itself, favorably affect the abnormal humoral immune state.

In addition to blocking hormone synthesis, propylthiouracil inhibits the peripheral deiodination of thyroxine to triiodothyronine. Methimazole does not have this effect and can antagonize the inhibitory effect of propylthiouracil. Although the quantitative significance of this inhibition has not been established, it does provide a theoretical rationale for the choice of propylthiouracil over other antithyroid drugs in the treatment of severe hyperthyroid states or of thyroid storm. In this acute situation, a decreased rate of conversion of circulating thyroxine to triiodothyronine would be beneficial.

Absorption, Metabolism, and Excretion. The antithyroid compounds currently used in the United States are propylthiouracil (6-n-propylthiouracil) and methimazole (1-methyl-2-mercaptoimidazole; TAPAZOLE). In Great Britian and Europe, carbimazole (NEO-MERCAZOLE), a carbethoxy derivative of methimazole, is available, and its antithyroid action is due to its conversion to methimazole after absorption. Some pharmacological properties of propylthiouracil and methimazole are shown in Table 56-5. Measurements of the course of organification of radioactive iodine by the thyroid show that absorption of effective amounts of propylthiouracil follows within 20 to 30 minutes of an oral dose. They also show that the duration of action of the compounds used clinically is brief. The effect of a dose of 100 mg of propylthiouracil begins to wane in 2 to 3 hours, and even a 500-mg dose is completely inhibitory for only 6 to 8 hours. As little as 0.5 mg of methimazole similarly decreases the organification of radioactive iodine in the thyroid gland, but a single dose of 10 to 25 mg is needed to extend the inhibition to 24 The half-life of propylthiouracil in plasma is about 75 minutes, whereas that for methimazole is 4 to 6 hours. The drugs appear to be concentrated in the thyroid, and methimazole, derived from the metabolism of carbimazole, accumulates after carbimazole is administered. Drugs and metabolites appear largely in the urine.

Although both propylthiouracil and methimazole cross the placenta and also can be found in milk, methimazole does so to a greater degree than propylthiouracil (Marchant *et al.*, 1977). The use of these drugs during pregnancy is discussed below.

Untoward Reactions. The incidence of side effects from propylthiouracil and methimazole as currently used is relatively low. The overall incidence as compiled from published cases by early investigators was 3% for propylthiouracil and 7% for methimazole, with 0.44% and 0.12% of cases, respectively, developing the most serious reaction, agranulocytosis (Meyer-Gessner et al., 1994). The development of agranulocytosis with methimazole may be dose-related, but no such relationship exists with propylthiouracil. Further observations have found little, if any, difference in side effects between these two agents, and suggest that an incidence of agranulocytosis of approximately 1 in 500 is a maximal figure. Agranulocytosis usually occurs during the first few weeks or months of therapy but may occur later. Because agranulocytosis can develop rapidly, periodic white-cell counts usually are of little help. Patients should immediately report the development of sore throat or fever, which usually heralds the onset of this reaction. Agranulocytosis is reversible upon discontinuation of the offending drug, and the administration of recombinant human granulocyte colony-stimulating factor may hasten recovery (Magner et al., 1994). Mild granulocytopenia, if noted, may be due to thyrotoxicosis

Table 56-5
Selected Pharmacokinetic Features of Antithyroid Drugs

	PROPYLTHIOURACIL	METHIMAZOLE
Plasma protein binding	~75%	Nil
Plasma half-life	75 minutes	~4-6 hours
Volume of distribution	~20 liters	~40 liters
Metabolism of drug during		
illness		***
Severe liver disease	Normal	Decreased
Severe kidney disease	Normal	Normal
Transplacental passage	Low	Increased
Levels in breast milk	Low	Increased

SOURCE: Adapted from Cooper, 1991b.

or may be the first sign of this dangerous drug reaction. Caution and frequent leukocyte counts are then required.

The most common reaction is a mild, occasionally purpuric, urticarial papular rash. It often subsides spontaneously without interrupting treatment, but it sometimes calls for the administration of an antihistamine or changing to another drug, because cross-sensitivity is uncommon. Other less frequent complications are pain and stiffness in the joints, paresthesias, headache, nausea, skin pigmentation, and loss of hair. Drug fever, hepatitis, and nephritis are rare, although abnormal liver function tests are not infrequent with higher doses of propylthiouracil.

Therapeutic Uses. The antithyroid drugs are used in the treatment of hyperthyroidism in the following three ways: (1) as definitive treatment, to control the disorder in anticipation of a spontaneous remission in Graves' disease; (2) in conjunction with radioactive iodine, to hasten recovery while awaiting the effects of radiation; and (3) to control the disorder in preparation for surgical treatment. There is no uniformity of opinion as to which form of treatment is the most desirable (Soloman et al., 1990), and this is often influenced by a variety of considerations, as discussed below.

The usual starting dose for propylthiouracil is 100 mg every 8 hours or 150 mg every 12 hours. When doses larger than 300 mg daily are needed, further subdivision of the time of administration to

ry 4 to 6 hours is occasionally helpful. Methimazole is effective an given as a single daily dose because of its relatively long plasma and intrathyroidal half-life, as well as its long duration of action. Failures of response to daily treatment with 300 to 400 mg of propylthiouracil or 30 to 40 mg of methimazole are most commonly due to noncompliance. Delayed responses also are noted in patients with very large goiters or those in whom iodine in any form has been given beforehand. Once euthyroidism is achieved, usually within 12 weeks, the dose of antithyroid drug can be reduced.

Response to Treatment. Hyperthyroidism may be of two kinds-Graves' disease and hyperthyroidism from one or more hyperfunctioning thyroid nodules; whichever the cause, the hyperthyroidism seems to respond to antithyroid drugs in the same way. After treatment is instituted, there is usually a latent period of several weeks before improvement is clearly manifest. In patients with large goiters and particularly if they are nodular, the response may be slower. The rate of response is determined by the quantity of stored hormone, the rate of turnover of hormone in the thyroid, the half-life of the. hormone in the periphery, and the completeness of the block in synthesis imposed by the dosage given. When large doses are continued, and sometimes with the usual dose, hypothyroidism may develop as a result of overtreatment. The earliest signs of hypothyroidism call for a reduction in dose; if by chance they have advanced to the point of discomfort, thyroid hormone can be given to hasten recovery. A full dose of levothyroxine can be given. The lower maintenance dose of antithyroid drug discussed above is instituted for continued therapy. Concomitant use of levothyroxine therapy along with antithyroid drugs has been reported to increase rates of remission of Graves' disease in Japan (Hashizume et al., 1991). However, this may represent differences in patient population, as well as the higher iodine intr'- in Japan.

After treatment is initiated, patients should be examined and thyrou function tests (serum free thyroxine index and total triiodothy-

ronine concentrations) measured every 2 to 4 months. Once euthyroidism is established, follow-up every 4 to 6 months is reasonable.

Control of the hyperthyroidism usually is associated with a decrease in goiter size, but if the thyroid enlarges, hypothyroidism probably has been induced. When this occurs, the new enlargement is quickly reversed by giving thyroid hormone. The presumption is, therefore, that TSH is secreted in excessive amounts in response to the hypothyroidism and can be suppressed by thyroid hormone.

Remissions. The antithyroid drugs have been used in many patients to control the hyperthyroidism of Graves' disease until a remission occurs. Early investigators reported that 50% of patients so treated for 1 year remained well without further therapy for long periods, perhaps indefinitely. More recent reports have indicated that a much smaller percentage of patients sustain remissions after such treatment. Increased dietary iodine has been implicated in the latter, less favorable rates.

Unfortunately, there is no way of predicting before treatment is begun which patients will eventually achieve a lasting remission and who will relapse. It is clear that a favorable outcome is unlikely when the disorder is of long standing, the thyroid is quite large, and various forms of treatment have failed. To complicate the issue further, it is thought that remission and eventual hypothyroidism may represent the natural history of Graves' disease.

During treatment, a fairly certain sign that a remission may have taken place is a reduction in the size of the goiter. The persistence of goiter usually indicates failure, unless the patient becomes hypothyroid. Another favorable indication is continued freedom from all signs of hyperthyroidism when the maintenance dose is small. Finally, a decrease in thyroid-stimulating immunoglobulins, suppression of ¹²³I thyroid uptake when thyroxine or triiodothyronine is given, and a normal serum TSH response to TRH are helpful in predicting a remission in some patients, although these tests are not routinely carried out.

The Therapeutic Choice. Because antithyroid drug therapy, radioactive iodine, and subtotal thyroidectomy all are effective treatments for Graves' disease, there is no worldwide consensus among endocrinologists as to the best approach to therapy. Two recent reviews have discussed available options (Franklyn, 1994, Klein et al., 1994). Prolonged drug therapy of Graves' disease in anticipation of a remission is most successful in patients with small goiters or mild hyperthyroidism. Those with large goiters or severe disease usually require definitive therapy with either surgery or radioactive iodine (131). Radioactive iodine remains the treatment of choice of many endocrinologists in the United States (Soloman et al., 1990). A relative contraindication for radioactive iodine therapy is coexisting, severe ophthalmopathy, since worsening of ophthalmopathy has been reported after radioactive iodine (Tallstedt et al., 1992; Kung et al., 1994). Depleting the thyroid gland of preformed hormone by treatment with antithyroid drugs is advisable in older patients prior to therapy with radioactive iodine so as to prevent a severe exacerbation of the hyperthyroid state during the subsequent development of radiation thyroiditis. Subtotal thyroidectomy is advocated for Graves' disease in young patients with large goiters, children who are allergic to antithyroid drugs, pregnant women (usually in the second trimester) who are allergic to antithyroid drugs, and patients who prefer surgery over antithyroid drugs or radioactive iodine. Radioactive iodine or surgery is indicated for definitive therapy in toxic nodular goiter, since remissions following antithyroid drug therapy do not occur.

Thyrotoxicosis in Pregnancy. Thyrotoxicosis occurs in about 0.2% of pregnancies and is caused most frequently by Graves' disease. Antithyroid drugs are the treatment of choice; radioactive iodine is clearly contraindicated (Momotani et al., 1986). Propylthiouracil is preferred over methimazole because of its lower transplacental passage (Marchant et al., 1977). Propylthiouracil dosage should be minimized to keep the serum free thyroxine index in the upper half of the normal range or slightly elevated. As pregnancy progresses, Graves' disease often improves. Indeed, it is not uncommon for patients to require daily propylthiouracil doses of less than 100 mg or to have antithyroid drugs discontinued by the end of pregnancy. Therefore, the propylthiouracil dose should be reduced, and maternal thyroid function should be monitored frequently to decrease chances of fetal hypothyroidism. Relapse or worsening of Graves' disease is common after delivery, and patients should be monitored closely. Propylthiouracil is the drug of choice in nursing women; the very small amounts of the drug that appear in breast milk do not appear to affect thyroid function in the suckling baby. A review of antithyroid drug therapy in pregnancy has been published recently (Mandel et al., 1994).

Adjuvant Therapy. Several drugs that have no intrinsic antithyroid activity are useful in the symptomatic treatment of thyrotoxicosis. β-Adrenergic receptor antagonists (Chapter 10) are effective in antagonizing the catecholaminergic effects of thyrotoxicosis by reducing the tachycardia, tremor, and stare and relieving palpitations, anxiety, and tension. Either propranolol, 20 to 40 mg four times daily, or atenolol, 50 to 100 mg daily, is usually given initially. Propranolol and esmolol can be given intravenously if needed. Propranolol, in addition to its β -adrenergic receptor antagonist action, has weak inhibitory effects on peripheral conversion of thyroxine to triiodothyronine. Ca2+ channel blockers (diltiazem, 60 to 120 mg four times daily) can be used to control tachycardia and decrease the incidence of supraventricular tachyarthythmias (see Chapter 35). These drugs should be discontinued once the patient is euthyroid.

Other drugs that are useful in the rapid treatment of the severely thyrotoxic patient are agents that inhibit the peripheral conversion of thyroxine to triiodothyronine. Dexamethasone (0.5 to 1 mg two to four times daily) and the iodinated radiological contrast agents, iopanoic acid (TELEPAQUE, 500 to 1000 mg once daily) and sodium ipodate (ORAGRAFIN, 500 to 1000 mg once daily) are effective in the short term but should not be used chronically.

Preoperative Preparation. Patients must be rendered euthyroid prior to subtotal thyroidectomy as definitive treatment for hyperthyroidism to reduce operative morbidity and mortality. It is possible to bring virtually 100% of patients to a euthyroid state; the operative mortality in these patients in the hands of an experienced thyroid surgeon is extremely low. Prior treatment with antithyroid drugs usually is successful in rendering the patient enthyroid for surgery. Iodide is added to the regimen for 7 to 10 days prior to surgery to decrease the vascularity of the gland, making it less friable and decreasing the difficulties for the surgeon. In the patient who is either allergic to antithyroid drugs or is noncompliant, a euthyroid state usually can be achieved by treatment with iopanoic acid, dexamethasone, and propranolol for 5 to 7 days prior to surgery. All of these drugs should be discontinued after surgery.

Thyroid Storm. Thyroid storm is an uncommon but life-threatening complication of thyrotoxicosis in which a severe form of the disease is usually precipitated by an intercurrent medical problem (Smallridge, 1992; Gavin, 1991). It occurs in untreated or partially treated thyrotoxic patients. Precipitating factors associated with thyrotoxic crisis include: infections, stress, trauma, thyroidal or nonthyroidal surgery, diabetic ketoacidosis, labor, heart disease, and radioactive iodine treatment.

Clinical features are similar to those of thyrotoxicosis, but more exaggerated. Cardinal features include fever (temperature usually over 38.5°C) and tachycardia out of proportion to the fever. Nausea, vomiting, diarrhea, agitation, and confusion are frequent presentations. Coma and death may ensue in up to 20% of patients. Thyroid function abnormalities are similar to those found in uncomplicated hyperthyroidism. Therefore, thyroid storm is primarily a clinical diagnosis.

Treatment includes supportive measures such as intravenous fluids, antipyretics, cooling blankets, and sedation. Antithyroid drugs are given in large doses. Propylthiouracil is preferred over methimazole because of its additional action of impairing peripheral conversion of thyroxine to triiodothyronine. The recommended initial dose of propylthiouracil is 200 to 300 mg every 6 hours. Propylthiouracil and methimazole can be administered by nasogastric tube or rectally if necessary. Neither of these preparations is available for parenteral administration in the United States.

Iodides, orally or intravenously, are used after the first dose of an antithyroid drug has been administered (see below). The radiographic contrast dyes may be used to block thyroid hormone release (as a result of the iodide released from these agents) and to inhibit thyroxine to triiodothyronine conversion. β -Adrenergic receptor antagonists, such as propranolol and esmolol, and Ca2+ channel blockers may also be used to control tachyarrhythmias. Dexamethasone (0.5 to 1 mg intravenously every 6 hours) is recommended both as supportive therapy and as an inhibitor of conversion of thyroxine to triiodothyronine. Finally, treatment of the underlying precipitating illness is essential.

Ionic Inhibitors

The term ionic inhibitors designates the substances that interfere with the concentration of iodide by the thyroid gland. The effective agents are themselves anions that in some ways resemble iodide; they are all monovalent, hydrated anions of a size similar to that of iodide. The most studied example, thiocyanate, differs from the others qualitatively; it is not concentrated by the thyroid gland, and in large amounts it inhibits the organification of iodine. Thiocyanate is produced following the enzymatic hydrolysis of certain plant glycosides. Thus, certain foods (e.g., cabbage) and cigarette smoking result in an increased concentration of thiocyanate in the blood and urine, as does the administration of sodium nitroprusside. Dietary precursors of thiocyanate may be a contributing factor in endemic goiter in certain parts of the world, especially in Central Africa, where the intake of iodine is very low (Delange et al., 1993).

Among other anions, perchlorate (ClO4") is ten times as active as thiocyanate. Although perchlorate can be used to control hyperthyroidism, it has caused fatal aplastic anemia when given in excessive amounts (2 to 3 g daily). Over the past few years, however, percholorate in doses of 750 mg daily has been used in the treatment of Graves' disease and amiodarone-induced thyrotoxicosis. Perchlorate can be used to "discharge" inorganic iodide from the thyroid gland in a diagnostic test of organification. Other ions, selected on the basis of their size, also have been found to be active; fluoborate (BF₄⁻) is as effective as perchlorate. Lithium has a multitude of effects on thyroid function; its principal effect is decreased secretion of thyroxine and triiodothyronine (Takami, 1994).

Iodide

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Iodide is the oldest remedy for disorders of the thyroid gland. Before the antithyroid drugs were used, it was the only substance available for control of the signs and symptoms of hyperthyroidism. Its use in this way is indeed paradoxical, and the explanation for this paradox is still incomplete.

Mechanism of Action. High concentrations of iodide appear to influence almost all important aspects of iodine metabolism by the thyroid gland (see Ingbar, 1972). The capacity of iodide to limit its own transport has been mentioned above. Acute inhibition of the synthesis of iodotyrosines and iodothyronines by iodide also is well known (the Wolff-Chaikoff effect). This transient, 2-day inhibition is observed only above critical concentrations of intracellular rather than extracellular concentration of iodide. With time there is "escape" from this inhibition that is associated with an adaptive decrease in iodide transport and a lowered intracellular iodide concentration (Braverman and Ingbar, 1963). The mechanism of the Wolff-Chaikoff effect may involve inhibition of inositol phoshate signaling pathways within the thyrocyte (Corvilain et al., 1994).

A very important clinical effect of high plasma iodide concentration is an inhibition of the release of thyroid hormone. This action is rapid and efficacious in severe thyrotoxicosis. The effect is exerted directly on the thyroid gland, and it can be demonstrated in the euthyroid subject and experimental animals as well as in the hyperthyroid patient. Recent studies in a cultured thyroid cell line suggest that some of the inhibitory effects of iodide on thyrocyte proliferation may be mediated by actions of iodide on crucial regulatory points in the cell cycle (Smerdely et al., 1993).

In euthyroid individuals, the administration of doses of iodide from 1.5 to 150 mg daily results in small decreases in plasma thyroxine and triiodothyronine concentrations and small compensatory increases in scrum TSH values, with all values remaining in the normal range. However, euthyroid patients with a history of a wide variety of underlying thyroid disorders may develop iodine-induced hypothyroidism when exposed to large amounts of iodine present in many commonly prescribed drugs (Table 56–6), and these patients do not escape from the acute Wolff-Chaikoff effect (Braverman, 1994). Among the disorders that predispose patients to iodine-induced hypothyroidism are: treated Graves' disease, Hashimoto's thyroiditis, postpartum lymphocytic thyroiditis, subacute painful thyroiditis, and lobectomy for benign nodules. The most commonly rescribed iodine-containing drugs are certain expectorants, topical antiseptics, and radiology contrast agents.

Response to Iodide in Hyperthyroidism. The response to iodides in patients with hyperthyroidism is often striking and rapid. The effect is usually discernible within 24 hours, and the basal metabolic rate may fall at a rate comparable to that following thyroidectomy. This provides evidence that the release of hormone into the circulation is rapidly blocked. Furthermore, thyroid hormone synthesis also may be decreased. The maximal effect is attained after 10 to 15 days of continuous therapy, when the signs and symptoms of hyperthyroidism may have greatly improved.

The changes in the thyroid gland have been studied in detail; vascularity is reduced, the gland becomes much firmer, the cells become smaller, colloid reaccumulates in the follicles, and the quantity of bound iodine increases. The changes are those that would be expected if the excessive stimulus to the gland had somehow been removed or antagonized.

Unfortunately, iodide therapy usually does not completely control the manifestations of hyperthyroidism, and after a variable period of time, the beneficial effect disappears. With continued treatment, the hyperthyroidism may return in its initial intensity or may become even more severe than it was at first. It is for this reason that, when iodide was the only agent available for the treatment of hyperthyroidism, it use was usually restricted to preparation of the patient for thyroidectomy.

Therapeutic Uses. The uses of iodide in the treatment of hyperthyroidism are in the preoperative period in preparation for thyroidectomy and, in conjunction with antithyroid drugs and propranolol, in the treatment of thyrotoxic crisis. Prior to surgery, iodide is sometimes employed alone, but more frequently it is used after the hyperthyroidism has been controlled by an antithyroid drug. It is then given during the 7 to 10 days immediately preceding the operation. Optimal control of hyperthyroidism is achieved if antithyroid drugs are first given alone. If iodine also is given from the beginning, variable responses are observed; sometimes the effect of iodide predominates, storage of hormone is promoted, and prolonged antithyroid treatment is required before the hyperthyroidism is controlled. These clinical observations may be explained by the ability of iodide to prevent the inactivation of thyroid peroxidase by antithyroid drugs (Taurog, 1991).

Another use of iodine is to protect the thyroid from radioactive iodine fallout following a nuclear accident. Because the uptake of radioactive iodine is inversely proportional to the serum concentration of stable iodine, the administration of 30 to 100 mg of iodide daily will markedly decrease the thyroid uptake of radioisotopes of iodine. Following the Chernobyl nuclear reactor accident in 1986, approximately 10 million children and adults in Poland were given stable iodide to block the thyroid exposure to radioactive iodine from the atmosphere and from dairy products from cows that ate contaminated grass (Naumann and Wolf, 1993).

The dosage or form in which iodide is administered bears little relationship to the response achieved in hyperthyroidism, provided not less than the minimal effective amount is given; this dosage is

Table 56-6
Commonly Used Iodine-Containing Drugs

DRUGS	IODINE CONTENT
Oral or local	
Amiodarone	75 mg/tablet
Calcium iodide (e.g., CALCIDRINE SYRUP)	26 mg/ml
Iodoquinol (diiodohydroxyquin)	134–416 mg/tablet
Echothiophate iodide ophthalmic solution	5–41 μ g/drop
Hydriodic acid syrup	13-15 mg/ml
Iodochlohydroxyquin	104 mg/tablet
Iodine-containing vitamins	0.15 mg/tablet
Iodinated glycerol	15 mg/tablet
Idoxuridine ophthalmic solution	18 μg/drop
Kelp	0.15 mg/tablet
Potassium iodide (e.g., QUADRINAL)	145 mg/tablet
Lugol's solution	6.3 mg/drop
Niacinamide hydroiodide + potassium iodide	3 1
(e.g., IODO-NIACIN)	115 mg/tablet
PONARIS nasal emollient	5 mg/0.8 ml
Saturated solution of potassium iodide	38 mg/drop
Parenteral preparations	o and and
Sodium iodide, 10% solution	85 mg/ml
Topical antiseptics	
Iodoquinol (diiodohydroxyquin) cream	6 mg/g
Iodine tincture	40 mg/ml
Iodochlorhydroxyquin cream	. 12 mg/g
Iodoform gauze	4.8 mg/100 mg gauze
Povidone iodine	10 mg/ml
Radiology contrast agents	
Diatrizoate meglumine sodium	370 mg/ml
Propyliodone	340 mg/ml
Iopanoic_acid	333 mg/tablet
Ipodate	308 mg/capsule
Tatholomoto	480 mg/ml
Tomaramare	483 mg/ml before dilution
Metrizamide The state of the latter of the state of the s	"" "AX3 mg/ml before dillition

SOURCE: Adapted from Braverman, 1994.

6 mg per day in most, but not all, patients. Strong todine solution (Lugol's solution) is widely used and consists of 5% iodine and 10% potassium iodide, which yields a dose of 6.3 mg of iodine per drop. The iodine is reduced to iodide in the intestine before absorption. Saturated solution of potassium iodide also is available, containing 38 mg per drop. Typical doses include 3 to 5 drops of Lugol's solution or 1 to 3 drops of saturated solution of potassium iodide 3 times a day. These doses have been determined empirically and are far in excess of that needed.

Untoward Reactions. Occasional individuals show marked sensitivity to iodide or to organic preparations that contain iodine when they are administered intravenously. The onset of an acute reaction

may occur immediately or several hours after administration. Angioedema is the outstanding symptom, and swelling of the larynx may lead to suffocation. Multiple cutaneous hemorrhages may be present. Also, manifestations of the serum-sickness type of hypersensitivity, such as fever, arthralgia, lymph node enlargement, and eosinophilia, may appear. Thrombotic thrombocytopenic purpura and fatal periarteritis nodosa attributed to hypersensitivity to iodide have also been described.

The severity of symptoms of chronic intoxication with iodide (iodism) is related to the dose. The symptoms start with an unpleasant brassy taste and burning in the mouth and throat, as well as soreness of the teeth and gums. Increased salivation is noted. Coryza, sneezing, and irritation of the eyes with swelling of the eyelids are

commonly observed. Mild iodism simulates a "head cold." The patient often complains of a severe headache that originates in the frontal sinuses. Irritation of the mucous glands of the respiratory tract causes a productive cough. Excess transudation into the bronchial tree may lead to pulmonary edema. In addition, the parotid and submaxillary glands may become enlarged and tender, and the syndrome may be mistaken for mumps parotitis. There also may be inflammation of the pharynx, larynx, and tonsils. Skin lesions are common, and vary in type and intensity. They usually are mildly acneform and distributed in the seborrheic areas. Rarely, severe and sometimes fatal eruptions (ioderma) may occur after the prolonged use of iodides. The lesions are bizarre, resemble those caused by bromism, a rare problem, and, as a rule, involute quickly when iodide is withdrawn. Symptoms of gastric irritation are common; and diarrhea, which is sometimes bloody, may occur. Fever is occasionally observed, and anorexia and depression may be present. The mechanisms involved in the production of these derangements remain unknown.

Fortunately, the symptoms of iodism disappear spontaneously within a few days after stopping the administration of iodide. The renal excretion of I⁻ can be increased by procedures that promote Cl⁻ excretion (e.g., osmotic diuresis, chloruretic diuretics, and salt loading). These procedures may be useful when the symptoms of iodism are severe.

Radioactive Iodine

Chemical and Physical Properties. Although iodine has several radioactive isotopes, greatest use has been made of 131 I. It has a half-life of 8 days, and, therefore, over 99% of its radiation is expended within 56 days. Its radioactive emissions include both γ rays and β particles. The short-lived radionuclide of iodine, 123 I, is primarily a γ -emitter with a half-life of only 13 hours. This permits a relatively brief exposure to radiation during thyroid scans.

Effects on the Thyroid Gland. The chemical behavior of the radioactive isotopes of iodine is identical to that of the stable isotope, 1271. 1311 is rapidly and efficiently trapped by the thyroid, incorporated into the iodoamino. acids, and deposited in the colloid of the follicles, from which it is slowly liberated. Thus, the destructive β particles originate within the follicle and act almost exclusively upon the parenchymal cells of the thyroid with little or no damage to surrounding tissue. The γ radiation passes through the tissue and can be quantified by external detection. The effects of the radiation depend upon the dosage. When small tracer doses of 131 are administered, thyroid function is not disturbed. However, when large amounts of radioactive iodine gain access to the gland, the characteristic cytotoxic actions of ionizing radiation are observed. Pyknosis and necrosis of the follicular cells are ollowed by disappearance of colloid and fibrosis of the gland. With properly selected doses of ¹³¹I, it is possible to destroy the thyroid gland completely without detectable

injury to adjacent tissues. After smaller doses, some of the follicles, usually in the periphery of the gland, retain their function.

Therapeutic Uses. Sodium iodide I 131 (IODOTOPE THERAPEUTIC) is available as a solution or in capsules containing essentially carrier-free ¹³¹I suitable for oral administration. Sodium iodide I 123 is available for scanning procedures. Radioactive iodine finds its widest use in the treatment of hyperthyroidism and in the diagnosis of disorders of thyroid function. Discussion will be limited to the uses of ¹³¹I.

Hyperthyroidism. Radioactive iodine is highly useful in the treatment of hyperthyroidism, and in many circumstances it is regarded as the therapeutic procedure of choice for this condition (Soloman et al., 1990; for review, see Farrar and Toff, 1991). The use of iodide as treatment for hyperthyroidism, however, may preclude, for months, treatment and certain imaging studies with radioactive iodine.

Desage and Technique. 1311 is administered orally, and the effective indicative iodine.

Dosage and Technique. 131I is administered orally, and the effective dose differs for individual patients. It depends primarily upon the size of the thyroid, the iodine uptake of the gland, and the rate of release of radioactive iodine from the gland subsequent to its deposition in the colloid. To determine these variables insofar as possible, many investigators administer a tracer dose of ¹³¹I and calcu-. late the ¹³¹I accumulated by the gland and the rate of loss therefrom. The weight of the gland is estimated by palpation. From these data, the dose of isotope necessary to provide from 7000 to 10,000 rad per gram of thyroid tissue is determined. Even when dosage is controlled in this manner, it is difficult to predict the response of an individual to a given amount of the isotope. For these reasons, the optimal dose of 131I, expressed in terms of microcuries taken up per gram of thyroid tissue, varies in different laboratories from 80 to 150 μCi. The usual total dose is 4 to 15 mCi. Lower-dosage ¹³¹I therapy (80 μ Ci/g thyroid) has been advocated to reduce the incidence of subsequent hypothyroidism. While the incidence of hypothyroidism in the early years after such therapy is lower, many patients with late hypothyroidism may go undetected, and the ultimate incidence of hypothyroidism is probably no less than with the larger doses (Glennon et al., 1972). In addition, relapse of the hyperthyroid state, or initial failure to alleviate the hyperthyroid state, is increased in patients receiving lower doses of [3] L. American view as a configurative

Course of Disease. The course of hyperthyroidism in a patient who has received an optimal dose of ¹³¹I is characterized by progressive recovery. It is very unusual for any tenderness to be noted in the thyroid region, and most observers have failed to detect any exacerbation of hyperthyroidism from loss of hormone from the damaged gland in patients whose preformed hormone stores have been depleted by antithyroid drug therapy. Beginning a few weeks after treatment, the symptoms of hyperthyroidism gradually abate over a period of 2 to 3 months. If therapy has been inadequate, the necessity for further treatment is apparent within 6 to 12 months.

Depending to some extent upon the dosage schedule adopted, one-half to two-thirds of patients are cured by a single dose, one-third to one-fifth require two doses, and the remainder require three or more doses before the disorder is controlled. Patients treated with larger doses of ¹³¹I almost always develop hypothyroidism within a few months.

Propranolol or antithyroid drugs or both can be used to hasten the control of hyperthyroidism while awaiting the full effects of the radioactive iodine. However, the antithyroid drugs should be withheld for a few days before and after the therapeutic dose of ¹³¹I.

Advantages. The advantages of radioactive iodine in the treatment of Graves' disease are many. No death as a direct result of the use of the isotope has been reported, and only by a gross miscalculation of dose could such an event conceivably occur. In the non-pregnant patient, no tissue other than the thyroid is exposed to sufficient ionizing radiation to be detectably altered. Nevertheless, the continuing concern about potential effects of radiation on germ cells prompts some endocrinologists to advocate antithyroid drugs or surgery in younger patients who are acceptable operative risks (Cooper, 1991b). Hypoparathyroidism is a small risk of surgery. With radioactive iodine treatment, the patient is spared the risks and discomfort of surgery. The cost is low, hospitalization is not required, and patients can indulge in their customary activities during the entire procedure.

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Disadvantages. The chief disadvantage of the use of radioactive iodine is the high incidence of delayed hypothyroidism that is induced. Even when elaborate procedures are used to estimate iodine uptake and gland size, a certain percentage of patients will be overtreated. A distressing feature of this complication is its rising prevalence with the passage of time; the longer the interval after treatment, the higher the incidence. Several analyses of groups of patients treated 10 or more years previously suggest that the eventual rate may exceed 80%. However, it now appears that the incidence of hypothyroidism also increases progressively after subtotal thyroidectomy, and such failure of glandular function is probably part of the natural progression of Graves' disease, no matter what the therapy.

Although it is often said that hypothyroidism is not a serious complication because it can be treated so easily with thyroid hormone, its onset may be insidious and overlooked for some time. Also, once diagnosed it is difficult to ensure that patients who need the hormone actually take it. Hypothyroidism is obviously a serious complication deserving of painstaking care to make certain that optimal replacement therapy is provided.

Another disadvantage of radioactive iodine therapy is the long period of time that is sometimes required before the hyperthyroidism is controlled. When a single dose is effective, the response is most satisfactory; however, when multiple doses are needed, it may be many months or a year or more before the patient is well. This disadvantage can be largely overcome if the initial dose is sufficiently large. Other disadvantages include possible worsening of ophthalmopathy after treatment, although this is controversial (Tallstedt et al., 1992). Although extremely rare, there have been reported cases of thyroid storm after therapy with ¹³¹I.

Indications. The clearest indication for this form of treatment is hyperthyroidism in older patients and in those with heart disease. Radioactive iodine also is the best form of treatment when Graves' disease has persisted or recurred after subtotal thyroidectomy and when prolonged treatment with antithyroid drugs has not led to remission. Finally, radioactive iodine is indicated in patients with toxic nodular goiter, since the disease does not go into spontaneous remission. The risk of inducing hypothyroidism is less in nodular goiter than in Graves' disease, perhaps because of the normal progression of the latter and the preservation of nonautonomous thyroid tissue in the former. Usually, larger doses of radioactive iodine are required in the treatment of toxic nodular goiter than in the treatment of Graves' disease.

Contraindications. The risk of causing neoplastic changes in the thyroid gland has been constantly under consideration since radioactive iodine was first introduced, and only small numbers of children have been treated in this way. Indeed, many clinics have declined to treat younger patients for fear of causing cancer and have reserved radioactive iodine for patients over some arbitrary age, such as 25 or 30 years. Since experience with ¹³¹I is now vast, these age limits are lower than they were in the past. There is no evidence that radioactive iodine therapy for Graves' disease has caused thyroid or any other form of cancer in adults, although the very large doses that are used to treat cancer (see below) may be associated with an increased incidence of leukemia. The use of radioactive iodine during pregnancy is contraindicated; after the first trimester the fetal thyroid would concentrate the isotope and thus suffer damage, but even during the first trimester radioactive iodine is best avoided because there may be adverse effects of radiation on fetal tissues.

Metastatic Thyroid Carcinoma. While most well-differentiated thyroid carcinomas accumulate very little iodine, stimulation of iodine uptake with TSH often is used effectively to treat metastases. Follicular carcinomas, which account for 10% to 15% of thyroid malignancies, are especially amenable to this treatment. Currently, endogenous TSH stimulation is evoked by withdrawal of thyroid hormone replacement therapy in patients previously treated with near-total thyroidectomy with or without radioactive ablation of residual thyroid tissue. In the future, injection of recombinant human TSH may be sufficient (Meier et al., 1994). Total body ¹³¹I scanning when the patient is hypothyroid (TSH > 35 mU/l), should be performed to identify metastatic disease or residual thyroid bed tissue. Depending upon the residual uptake, or the presence of metastatic disease, an ablative dose of ¹³¹I ranging from 30 to 150 mCi is administered, and a repeat total body scan is obtained 1 week later. The precise amount of 131I needed to treat residual tissue and metastases is controversial.

Suppressive therapy with levothyroxine is indicated in all patients after treatment for thyroid cancer. The goal of therapy is to keep serum TSH levels in the subnormal range (Burmeister et al., 1992). Follow-up evaluation every 6 months is reasonable, along with determination of serum thyroglobulin concentrations. A rise in serum thyroglobulin concentration is often the first indication of recurrent disease. Prognosis in patients with thyroid cancer depends upon the pathology and size of the turnor and is generally worse in the elderly (see Farid et al., 1994). Overall, the vast majority of patients with thyroid cancer will not die of their disease. Papillary cancer is not an aggressive tumor. It metastasizes locally and has a 10-year survival rate of greater than 90%. Lymph node metastases at the time. of diagnosis do little to alter the prognosis. Follicular cancer is more aggressive and can metastasize via the bloodstream. Still, prognosis is fair, and long-term survival is common. Anaplastic cancer is the exception, as it is highly malignant with survival usually less than 1 year.

Diagnostic Uses. Tracer studies with radioactive iodine have found wide application in studies of disorders of the thyroid gland. Measurement of the thyroidal accumulation of a tracer dose is helpful in the diagnosis of hyperthyroidism, hypothyroidism, and goiter, and the response of the thyroid to TSH or to suppression by thyroid hormone can be evaluated in this way. Following the administration of a tracer dose, the pattern of localization in the thyroid gland can be depicted by a special scanning apparatus, and this technique is sometimes useful in defining thyroid nodules as functional ("hot") or nonfunctional ("cold") and in finding ectopic thyroid tissue and occasionally metastatic thyroid tumors.

For further discussion of diseases of the thyroid, see Chapter 334 in Harrison's Principles of Internal Medicine, 13th ed., McGraw Hill, New York, 1994.

BIBLIOGRAPHY

- Astwood, E.B. Chemotherapy of hyperthyroidism. Harvey Lect., 1945, 40:195-235.
- Baran, D.T. Thyroid hormone and bone mass: The clinician's dilemma [editorial]. Thyroid, 1994, 4:143-144.
- Benevenga, S., Cahnmann, H.J., Rader, D., Kindt, M., and Robbins, J. Thyroxine binding to the apolipoproteins of high density lipoproteins HDL₂ and HDL₃. Endocrinology, 1992, 131:2805-2811.
- Berry, M.J., Banu, L., and Larsen, P.R. Type I iodothyronine 5'-deiodinase is a selenocystine-containing enzyme. *Nature*, 1991, 349:438-440.
- Berry, M.J., and Larsen, P.R. The role of selenium in thyroid hormone action. *Endocr. Rev.*, 1992, 13:207-219.
- Braverman, L.E., and Ingbar, S.H. Changes in thyroidal function during adaptation to large doses of iodine. J. Clin. Invest., 1963, 42:1216-1231.
- Brent, G.A., and Hershman, J.M. Thyroxine therapy in patients with severe nonthyroidal illnesses and low serum thyroxine concentration. J. Clin. Endocrinol. Metab., 1986, 63:1-8.
- Burmeister, L.A., Goumaz, M.O., Mariash, C.N., and Oppenheimer, J.H. Levothyroxine dose requirements for thyrotropin suppression in the treatment of differentiated thyroid cancer. *J. Clin. Endocrinol. Metab.*, 1992, 75:344–350.
- Chanoine, J.P., Braverman, L.E., Farwell, A.P., Safran, M., Alex, S., Dubord, S., Braverman, L.E., and Leonard, J.L. The thyroid gland is a major source of circulating T3 in the rat. J. Clin. Invest., 1993, 91:2709-2713.
- Corda, D., Marcocci, C., Kohn, L.D., Axelrod, J., and Luini, A. Association of the changes in cytosolic Ca²⁺ and iodide efflux induced by thyrotropin and by stimulation of alpha 1-adrenergic receptors in cultured rat thyroid cells. *J. Biol. Chem.*, 1985, 260:9230-9236.
- Corvilain, B., Laurent, E., Lecomte, M., Vansande, J., and Dumont, J.E. Role of the cyclic adenosine 3',5'-monophosphate and the phosphatidylinositol-Ca²⁺ cascades in mediating the effects of thyrotropin and iodide on hormone synthesis and secretion in human thyroid slices.—

 J. Clin. Endocrinol. Metab., 1994, 79:152-159.
- Cushing, H. The Pituitary Body and Its Disorders. J.B. Lippincon Co., Philadelphia, 1912.
- Davidson, B, Soodak, M, Neary, J.T., Strout, H.V., Kieffer, J.D., Mover, H., and Maloof, F. The irreversible inactivation of thyroid peroxidase by methylmercaptoimidazole, thiouracil and propylthiouracil in vitro and its relationship to in vivo findings. Endocrinology, 1978, 103:871-882.
- Davis, P.J., Davis, F.B., and Lawrence, W.D. Thyroid hormone regulation and membrane Ca²⁺-ATPase activity *Endocr. Res.*, 1989, 15:651-682.
- Dunn, A.D., Crutchfield, H.E., and Dunn, J.T. Proteolytic processing of thyroglobulin by extracts of thyroid lysosomes. *Endocrinology*, 1991, 128:3073-3080.
- Dunn, A.D., and Dunn, J.T. Cysteine proteinases from human thyroids and their actions on thyroglobulin. Endocrinology, 1988, 123:1089-1097.
- nn, J.T., Anderson, P.C., Fox, J.W., Fassler, C.A., Dunn, A.D., Hite, L.A., and Moore, R.C. The sites of thyroid hormone formation in rabbit thyroglobulin. *J. Biol. Chem.*, 1987, 262:16948–16952.

- Engler, H, Taurog, A., and Nakashima, T. Mechanism of inactivation of thyroid peroxidase by thioureylene drugs. *Biochem. Pharmacol.*, 1982, 31:3801-3806
- Everett, A.W., Umeda, P.K., Sinha, A.M., Rabinowitz, M., and Zak, R. Expression of myosin heavy chains during thyroid hormone-induced cardiac growth. Fed. Proc., 1986, 45:2568-2572.
- Farsetti, A., Mitsuhashi, T., Desvergne, B., Robbins, J., and Nikodem, V.M. Molecular basis of thyroid hormone regulation of myelin basic protein gene expression in rodent brain. J. Biol. Chem., 1991, 266:23226-23232.
- Farwell, A.P., Lynch, R.M., Okulicz, W.C., Comi, A.M., and Leonard, J.L. The actin cytoskeleton mediates the hormonally regulated translocation of type II iodothyronine 5'-deiodinase in astrocytes. J. Biol. Chem., 1990, 265:18546-18553.
- Field, J.B., Ealey, P.A., Marshall, N.J., and Cockcroft, S. Thyroid-stimulating hormone stimulates increases in inositol phosphates as well as cyclic AMP in the FRTL-5 rat thyroid cell line. *Biochem. J.*, 1987, 247:519-524.
- Glennon, J.A., Gordon, E.S., and Sawin, C.T. Hypothyroidism after low-dose ¹³¹I treatment of hyperthyroidism. Ann. Intern. Med., 1972, 76:721-723.
- Glinoer, D., Riahi, M., Grun, J.P., and Kinthaert, J. Risk of subclinical hypothyroidism in pregnant women with asymptomatic autoimmune thyroid disorders. J. Clin. Endocrinol. Metab., 1994, 79:197-204.
- Greer, M.A., Grimm, Y., and Studer, H. Qualitative changes in the secretion of thyroid hormones induced by iodine deficiency. *Endocrinology*, 1968, 83:1193-1198.
- Gross, G., Sykes, M., Arellano, R., Fong, B., and Angel, A. HDL clear-ance and receptor-mediated catabolism of LDL are reduced in hypothyroid rats. Atherosclerosis, 1987, 66:269-275.
- Gross, J., and Pitt-Rivers, R. The identification of 3:5:3'-1-triiodothyronine in human plasma. *Lancet*, 1952, 1:439-441.
- Gross, J., and Pitt-Rivers, R. 3:5:3'-Triiodothyronine. 1. Isolation from thyroid gland and synthesis. *Biochem. J.*, 1953a, 53:645-652. 2. Physiological activity, *Ibid.* 1953b, 53:652-657.
- Harington, C.R. Biochemical basis of thyroid function. Lancet, 1935, -1:1199-1204, 1261-1266.
- Hashizume, K., Ichikawa, K., Sakurai, A., Suzuki, S., Takeda, T., Kobayashi, M., Miyamoto, T., Arai, M., and Nagasawa, T. Administration of thyroxine in treated Graves' disease: effects on the level of antibodies to thyroid-stimulating hormone receptors and on the risk of recurrance of hyperthyroidism. N. Engl. J. Med., 1991, 324:947-953.
- Hays, M.T., Localization of human thyroxine absorption. Thyroid, 1991, 1:241-248.
- Hays, M.T., and Nielsen, K.R. Human thyroxine absorption: age effects and methodological analyses. *Thyroid*, 1994, 4:55-64.
- Jorgensen, E.C. Stereochemistry of thyroxine and analogues. Mayo Clin. Proc., 1964, 39:560-568.
- Kaplan, M.M. Assessment of thyroid function during pregnancy. Thyroid, 1992, 2:57-61.
- Kung, A.W., Yau, C.C., and Cheng, A. The incidence of ophthalmopathy after radioiodine therapy for Graves' disease: prognostic factors and

- the role of methimazole. J. Clin. Endocrinol. Metab., 1994, 79:542-546.
- Laurent, E., Mockel, J., Van Sande, J., Graff, I., and Dumont, J.E. Dual activation by thyrotropin of the phospholipase C and cyclic AMP cascades in human thyroid. Mol. Cell. Endocrinol., 1987, 52:273-278.
- Leeson, P.D., Emmett, J.C., Shah, V.P., Showell, G.A., Novelli, R., Prain, D., Benson, M.G., Ellis, D., Pearce, N.J., and Underwood, A.H. Selective thyromimetics. Cardiac-sparing thyroid hormone analogues containing 3'-arylmethyl substituents. J. Med. Chem., 1989, 32:320-336.
- Leonard, J.L., Kaplan, M.M., Visser, T.J., Silva, J.E., and Larsen, P.R. Cerebral cortex responds rapidly to thyroid hormones. Science, 1981, 214:571-573.
- Magner, J.A., and Synder, D.K. Methimazole-induced agranulocytosis treated with recombinant human granulocyte colony-stimulating factor (G-CSF). Thyroid, 1994, 4:295-296.
- Magnusson, R.P., Gestautas, J., Taurog, A., and Rapoport, B. Molecular cloning of the structural gene for porcine thyroid peroxidase. J. Biol. Chem., 1987, 262:13885-13888.
- Magnusson, R.P., Taurog, A., and Dorris, M.L. Mechanisms of thyroid peroxidase- and lactoperoxidase-catalyzed reactions involving iodide. J. Biol. Chem., 1984, 259:13783-13790.
- Manley, S.W., Rose, D.S., Huxham, G.J., and Bourke, J.R. Role of calcium in the secretomotor response of the thyroid: effects of calcium ionophore A23187 on radioiodine turnover, membrane potential and fluid transport in cultured porcine thyroid cells. J. Endocrinol., 1988, 116:373-380.
- Marchant, B., Brownlie, B.E.W., Hart, D.W., Horton, P.W., and Alexander, W.D. The placental transfer of propylthiouracil, methimazole and carbimazole. J. Clin. Endocrinol. Metab., 1977, 45:1187-1193.
- Marine, D., and Kimball, O.P. The prevention of simple goiter in man: a survey of the incidence and types of thyroid enlargements in the schoolgirls of Akron, Ohio, from the 5th to the 12th grades, inclusive; the plan of prevention proposed. J. Lab. Clin. Med., 1917, 3:40-48.
- McKinney, J.D., and Waller, C.L. Polychlorinated biphenyls as hormonally active structural analogues. Environ. Health Perspect., 1994, 102:290-297.
- Meier, C.A., Braverman, L.E., Ebner, S.A., Veronikis, I., Daniels, G.H., Ross, D.S., Deraska, D.J., Davies, T.F., Valentine, M., DeGroot, L.J., Curran, P., McEllin, K., Reynolds, J., Robbins, J., and Weintraub, B.D. Dingnostic use of recombinant human thyrotropin in patients with thyroid carcinoma (Phase VII study). J. Clin. Endocrinol. Metab., 1994, 28:188-196.
- Momotani, N., Noh, J., Oyanagi, H., Ishikawa, N., and Ito, K. Antithyroid drug therapy for Graves' disease during pregnancy. Optimal regimen for fetal thyroid status. N. Engl. J. Med., 1986, 315:24-28.
- Nauman, J., and Wolff, J. Iodide prophylaxis in Poland after the Chernobyl reactor accident: benefits and risks. Am. J. Med., 1993, 94:524-532.
- Nelson, J.C., and Tomei, R.T. Direct determination of free thyroxin in undiluted serum by equilibrium dialysis/radioimmunoassay. Clin. Chem., 1988, 34:1737-1744.
- Nelson, J.C., and Tomei, R.T. Dependence of the thyroxin/ thyroxin-binding globulin (TBG) ratio and the free thyroxin index on TBG concentrations. Clin. Chem., 1989, 35:541-544.
- Nelson, J.C., and Weiss, R.M. The effect of serum dilution on free thyroxine (T₄) concentration in the low T₄ syndrome of nonthyroidal illness. J. Clin. Endocrinol. Metab., 1985, 61:239-246.
- Nunez, J., and Correze, C. Interdependent effects of thyroid hormones and cAMP on lipolysis and lipogenesis in the fat cell. Adv. Cyclic Nucleotide Res., 1981, 14:539-554.

- Parmentier, M., Libert, F., Maenhaut, C., Lefort, A., Gerard, C., Perret, J., Van Sande, J. Dumont, J.E., and Vassart, G. Molecular cloning of the thyrotropin receptor. Science, 1989, 246:1620-1622.
- Roche, J., Lissitzky, S., and Michel, R. Sur la triiodothyronine, produit intermédiare de la transformation de la diiodothyronine en thyroxine. C. R. Acad. Sci. [D] (Paris), 1952a, 234:997-998.
- Roche, J., Lissitzky, S., and Michael, R. Sur la présence detriiodothyronine dans la thyroglobuline. C.R. Acad. Sci. [D](Paris), 1952b, 234:1228-1230.
- Rohrer, D., and Dillman, W.H. Thyroid hormone markedly increases the mRNA coding for sarcomeric reticulum Ca²⁺-ATPase in the rat heart. J. Biol. Chem., 1989, 263:6941-6944.
- Ros, M., Northup, J.K., and Malbon, C.C. Steady-state levels of G-proteins and β-adrenergic receptors in rat fat cells. Permissive effects of thyroid hormones. J. Biol. Chem., 1988, 263:4362-4368.
- Roti, E., Minelli, R., Gardini, E., and Braverman, L.E. The use and misuse of thyroid hormone. *Endocr. Rev.*, 1993, 14:401-423.
- Ruiz, M., Rajatanavin, R., Young, R.A., Taylor, C., Brown, R., Braverman, L.E., and Ingbar, S.H. Familial dysalbuminemic hyperthyroxinemia: a syndrome that can be confused with thyrotoxicosis. N. Engl. J. Med., 1982, 306:635-639.
- Safran, M., Farwell, A.P., and Leonard, J.L. Evidence that type II 5' deiodinase is not a selenoprotein. J. Biol. Chem., 1991, 266:13477-13480.
- Salter, A.M., Fisher, S.C., and Brindley, D.N. Interactions of triiodothyronine, insulin, and dexamethasone on the binding of human LDL to rat hepatocytes in monolayer culture. *Atherosclerosis*, 1988, 71:77-80.
- Samuels, H.H., Forman, B.M., Horowitz, Z.D., and Ye, Z.-S. Regulation of gene expression by thyroid hormone. J. Clin. Invest., 1988, 81:957-967.
- Sap, J., Munoz, A., Damm, K., Goldberg, Y., Ghysdael, J., Leutz, A., Beug, H., and Vennstrom, B. The c-erb-A protein is a high affinity receptor for thyroid hormone. Nature, 1986, 324:635-640.
- Scarabottolo, L., Trezzi, E., Roma, P., and Catapano, A.L. Experimental hypothyroidism modulates the expression of low density lipoprotein receptor by the liver. Atherosclerosis, 1986, 59:329-333.
- Sherman, S.I., and Ladenson, P.W. Organ-specific effects of tiratricol: a thyroid hormone analog with hepatic, not pituitary, superagonist effects. J. Clin. Endocrinol. Metab., 1992, 75:901-905.
- Simmonds, M. Ueber Hypophysisschwund mit todlichem Ausang. Disch. Med. Wochenschr., 1914, 40:322-323.
- Smerdely, P., Pitsiavas, V., and Boyages, S.C. Evidence that the inhibitory effects of iodide on thyroid cell proliferation are due to arrest of the cell cycle at GOG1 and G2M phases. *Endocrinology*, 1993, 133:2881-2888.
- Soloman, B., Glinoer, D., Lagasse, R., and Wartofsky, L. Current trends in the management of Graves' disease. J. Clin. Endocrinol. Metab. 1990, 70:1518-1524.
- Sterling, K. Direct triiodothyronine (T₃) action by a primary mitochondrial pathway. Endocr. Res., 1989, 15:683-715.
- Strait, K.A., Schwartz, H.L., Perez-Castillo, A., and Oppenheimer, J.H. Relationship of c-erbA mRNA content to tissue triiodothyronine nuclear binding capacity and function in developing and adult rats.

 J. Biol. Chem., 1990, 265:10514-10521.
- Takami, H. Lithium in the preoperative preparation of Graves' disease. Int. Surg., 1994, 79:89-90.
- Takasu, N., Yamada, T., and Shimizu, Y. Generation of H₂O₂ is regulated by cytoplasmic free calcium in cultured porcine thyroid cells. *Biochem. Biophys. Res. Commun.*, 1987, 148:1527-1532.
- Tallstedt, L., Lundell, G., Tørring, O., Wallin, G., Ljunggren, J.-G., Blomgren, H., Taube, A., and the Thyroid Study Group. Occurrence of ophthalmopathy after treatment for Graves' hyperthyroidism. N. Engl. J. Med., 1992, 326:1733-1738.

5 00 3

- Taurog, A. The mechanism of action of thioureylene antithyroid drugs. Endocrinology, 1976, 98:1031-1046.
- Thilly, C. H., Delange, F., Goldstein-Golaire, J., and Ermans, A.M. Endemic goiter prevention by iodized oil: a reassessment. J. Clin. Endocrinol. Metab., 1973, 36:1196-1204.
- Underwood, A.H., Emmett, J.C., Ellis, D., Flynn, S.B., Leeson, P.D., Benson, G.M., Novelli, R., Pearce, N.J., and Shah, V.P. A thyromimetic that decreases plasma cholesterol levels without increasing cardiac activity. *Nature*, 1986, 324:425-429.
- Van Sande, J., Raspe, E., Perret, J., Lejeune, C., Maenhaut, C., Vassart, G., and Dumont, J.E. Thyrotropin activates both the cyclic AMP and the PIP₂ cascades in CHO cells expressing the human cDNA of the TSH receptor. *Mol. Cell. Endocrinol.*, 1990, 74:R1-R6.
- Visser, T.J., Leonard, J.L., Kaplan, M.M., and Larsen, P.R. Kinetic evidence suggesting two mechanisms for iodothyronine 5'-deiodination in rat cerebral cortex. Proc. Natl. Acad. Sci. U.S.A., 1982, 79:5080-5084.
- Weinberger, C., Thompson, C.C., Ong, E.S., Lebo, R., Gruol, D.J., and Evans, R.M. The c-erb-A gene encodes a thyroid hormone receptor. *Nature*, 1986, 324:641-646.
- Wilke, T.J. Estimation of free thyroid hormone concentrations in the clinical laboratory. Clin. Chem., 1986, 32:585-592.

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- Bahn, R.S., and Heufelder, A.E. Pathogenesis of Graves' ophthalmopathy. N. Engl. J. Med., 1993, 329:1468-1475.
- Bottazzo, G.F., and Doniach, D. Autoimmune thyroid disease, Annu. Rev. Med., 1986, 37:353-359.
- 3raverman, L.E. Iodine and the thyroid: 33 years of study. *Thyroid*, 1994, 4:351-356.
- Braverman, L.E., Eber, O., and Langsteger, W. Heart and Thyroid. Black-well-MZV, Vienna, 1994.
- Braverman, L.E., and Refetoff, S. Clinical and Molecular Diseases of the Thyroid. Endocrine Society Press, Bethesda, 1994.
- Braverman, L.E., and Utiger, R.D. Weiner and Ingbar's The Thyroid.

 J.B. Lippincott Co., Philadelphia, 1991.
- Brent, G.A. The molecular basis of thyroid hormone action. N. Engl. J. Med., 1994, 331:847-853.
- Burch, H.B., and Wartofsky, L. Graves' ophthalmopathy: current concepts regarding pathogenesis and management. *Endocr. Rev.*, 1993, 14:747-793.
- Burman, K.D., and Baker, J.R., Jr. Immune mechanisms in Graves' disease. Endocr. Rev., 1985, 6:183-232.
- Cody, V. Thyroid hormone interactions: molecular conformation, protein binding and hormone action. *Endocr. Rev.*, 1980, 1:140-166.
- Cody, V. Thyroid hormone structure-function relationships. In, Werner and Ingbar's The Thyroid. (Braverman, L.E., and Utiger, R.D., eds.)

 J.B. Lippincott Co., Philadelphia, 1991, pp. 225-229.
- Cooper, D.S. Subclinical hypothyroidism. Adv. Endocrinol. Metab., 1991a, 2:77-89.
- Cooper, D.S. Treatment of thyrotoxicosis. In, Werner and Ingbar's The Thyroid. (Braverman, L.E., and Utiger, R.D., eds.) J.B. Lippincott Co., Philadelphia, 1991b, pp. 887-916.
- Cooper, D.S. Antithyroid drugs. N. Engl. J. Med., 1984, 311:1353-1362. Delange, F., Dunn, J.T., and Glinoer, D. Iodine Deficiency in Europe: A Continuing Concern. Plenum Press, New York, 1993.
- Dussault, J.H., and Ruel, J. Thyroid hormones and brain development.

 Annu. Rev. Physiol., 1987, 49:321-334.

- Farid, N.R., Shi, Y., and Zou, M. Molecular basis of thyroid cancer. Endocr. Rev., 1994, 15:202-232.
- Farrar, J.J., and Toft, A.D. Iodine-131 treatment of hyperthyroidism: current issues. Clin. Endocrinol., 1991, 35:207-212.
- Fisher, D.A. Management of congenital hypothyroidism. J. Clin. Endocrinol. Metab., 1991, 72:523-529.
- Franklyn, J.A. The management of hyperthyroidism. N. Engl. J. Med., 1994, 330:1731-1738.
- Gaitan, E. Environmental Goitrogenesis. CRC Press, Boca Raton, FL, 1989.
 Gammage, M.D., and Franklyn, J.A. Amiodarone and the thyroid. Q. J. Med., 1987, 62:83-86.
- Gavin, L.A. Thyroid crises. Med. Clin. North Am., 1991, 75:179-193.
 Gershengorn, M.C. Mechanism of thyrotropin releasing hormone stimulation of pituitary hormone secretion. Annu. Rev. Physiol., 1986, 48:515-526.
- Gharib, H., and Goellner, J.R. Fine-needle aspiration biopsy of the thyroid: an appraisal. *Ann. Intern. Med.*, 1993, 118:282-289.
- Glinoer, D. Maternal thyroid function in pregnancy. J. Endocrinol. Invest., 1993, 16:374-378.
- Gottlieb, P.A., and Braverman, L.E. The effect of thyroid disease on diabetes. Clin. Diabetes, 1994, 12:15-18.
- Green, W.L. Antithyroid compounds. In, Werner and Ingbar's The Thyroid. (Braverman, L.E., and Utiger, R.D., eds.) J.B. Lippincott Co., Philadelphia, 1991, pp. 322-334.
- Ingbar, S.H. Autoregulation of the thyroid. Response to iodide excess and depletion. Mayo Clin. Proc., 1972, 47:814-823.
- Kaptein, E.M. Thyroid hormone metabolism in illness. In, Thyroid Hormone Metabolism. Basic and Clinical Endocrinology, Vol. 8. (Hennemann, G., ed.) Marcel Dekker, Inc., New York, 1986, pp. 297-333.
- Klein, I., Becker, D.V., and Levey, G.S. Treatment of hyperthyroid disease. Ann. Intern. Med., 1994, 121:281-288.
- Larsen, P.R., Silva, J.E., and Kaplan, M.M. Relationships between circulating and intracellular thyroid hormones: physiological and clinical implications. *Endocr. Rev.*, 1981, 2:87-102.
- Lazar, M.A. Thyroid hormone receptors: multiple forms, multiple possibilities. *Endocr. Rev.*, 1993, 14:184-193.
- Legrand, J. Morphogenic actions of thyroid hormones. *Trends Neurosci.*, 1979, 2:234-236.
- Leonard, J.L., and Visser, T.J. Biochemistry of deiodination. In, Thyroid Hormone Metabolism. (Hennemann, G., ed.) Basic and Clinical Endocrinology, Vol. 8. Marcel Dekker, Inc., New York, 1986, pp. 189-230.
- Mandel, S.J., Brent, G.A., and Larsen, P.R. Review of antithyroid drug use during pregnancy and report of a case of aplasia cutis. *Thyroid*, 1994, 4:129-133.
- Mangelsdorf, D.J., Umesono, K., and Evans, R.M. The retinoid receptors. In, *The Retinoids: Biology, Chemistry, and Medicine*, 2nd ed. (Sporn, M.B., Roberts, A.B., and Goodman, D.S., eds.) Raven Press, New York, 1994, pp. 319-349.
- Mazzaferri, E.L. Management of a solitary thyroid nodule. N. Engl. J. Med. 1993, 328:553-559.
- McLachlan, S.M., and Rapoport, B. The molecular biology of thyroid peroxidase: cloning, expression and role as autoantigen in autoimmune thyroid disease. *Endocr. Rev.*, 1992, 13:192-206.
- Mendel, C.M. The free hormone hypothesis: a physiologically based mathematical model. *Endocr. Rev.*, 1989, 10:232-274.
- Meyer-Gessner, M., Benker, G., Lederbogen, S., Olbricht, T., and Reinwein, D. Antithyroid drug-induced agranulocytosis: clinical experience with ten patients treated at one institution and review of the literature. J. Endocrinol. Invest., 1994, 17:29-36.

- Nagayama, Y., and Rapoport, B. The thyrotropin receptor 25 years after its discovery: new insight after its molecular cloning. *Mol. Endocrinol.*, 1992, 6:145-156.
- Nicoloff, J.T., and Spencer, C.A. The use and misuse of the sensitive thyrotropin assays. J. Clin. Endocrinol. Metab., 1990, 71:553-558.
- Oppenheimer, J.H. Thyroid hormone action at the molecular level. In,

 Werner and Ingbar's The Thyroid. (Braverman, L.E., and Utiger, R.D.,
 eds.) J.B. Lippincott Co., Philadelphia, 1991, pp. 204-224.
- Oppenheirner, J.H., Schwartz, H.L., Mariash, C.N., Kinlaw, W.B., Wong, N.C.W., and Freake, H.C. Advances in our understanding of thyroid hormone action at the cellular level. *Endocr. Rev.*, 1987, 8:288-308.
- Porterfield, S.P., and Hendrich, C.E. The role of thyroid hormones in prenatal and neonatal neurological development: current perspectives. *Endocr. Rev.*, 1993, 14:94–106.
- Ross, D.R. Subclinical thyrotoxicosis. Adv. Endocrinol. Metab., 1991, 2:89-103.
- Roti, E., and Emerson, C.H. Postpartum thyroiditis. J. Clin. Endocrinol. Metab., 1992, 74:3-5.

- Siegrist-Kaiser, C.A., Juge-Aubry, C., Tranter, M.P., Ekenbarger, D.M., and Leonard, J.L. Thyroxine-dependent modulation of actin polymerization in cultured astrocytes. A novel, extranuclear action of thyroid hormone. J. Biol. Chem., 1990, 265:5296-5302.
- Smallridge, R.C. Metabolic and anatomic thyroid emergencies: a review. Crit. Care Med., 1992, 20:276-291.
- Surks, M.I., Chopra, I.J., Mariash, C.N., Nicoloff, J.T., and Solomon, D.H. American Thyroid Association guidelines for use of laboratory tests in thyroid disorders. J.A.M.A., 1990, 263:1529-1532.
- Taurog, A. Hormone synthesis: thyroid iodine metabolism. In, Werner and Ingbar's The Thyroid. (Braverman, L.E., and Utiger, R.D., eds.) J.B. Lippincott Co., Philadelphia, 1991, pp. 51-97.
- Toft, A.D. Thyroxine therapy. N. Engl. J. Med., 1994, 331:174-180.
- Van Etten, C.H. Goitrogens. In, Toxic Constituents of Plant Foodstuffs (Liener, I.E., ed.) Academic Press, Inc., New York, 1969, pp. 103-142.
- Vassart, G., and Dumont, J.E. The thyrotropin receptor and the regulation of thyrocyte function and growth. Endocr. Rev., 1992, 13:596-611.

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